

## Research Article

Ceylan Hepokur\*, Sema Misir, Tutku Tunç, Ugur Tutar, Ali Ihsan Hepokur and Mehmet Çiçek



# *In vitro* antimicrobial, antioxidant, cytotoxic activities, and wound healing potential of *Thymbra capitata* ethanolic extract

## [*Thymbra capitata* etanolik ekstraktının *in vitro* antimikrobiyal, antioksidan, sitotoksik aktiviteleri ve yara iyileşme potansiyeli]

<https://doi.org/10.1515/tjb-2019-0470>

Received November 25, 2019; accepted July 14, 2020;

published online December 14, 2020

### Abstract

**Objectives:** In this study, we aimed to detect the chemical compounds of *Thymbra capitata* ethanolic extract (TC-EtOH) as well as to evaluate its antimicrobial, antioxidant, cytotoxic activities, and *in vivo* wound healing effects.

**Methods:** The chemical composition of TC-EtOH was analyzed by Gas chromatography-mass spectrometry (GC-MS). Antioxidant and antimicrobial properties were determined with 2,2-diphenyl-1-picrylhydrazyl (DPPH), disc diffusion test and broth micro-dilution (minimal inhibitory concentration [MIC]) methods, respectively. Cytotoxic activity was tested on MG63 (human osteosarcoma) and MCF-7 (human breast carcinoma) cells by 2,3-bis-(2-methoxy-4-nitro-5-sulfophenyl)-2H-tetrazolium-

5-carboxanilide (XTT) assay. Tumor necrosis factor alpha (TNF- $\alpha$ ) protein levels were determined by ELISA.

**Results:** The major components of TC-EtOH were tetratriacontane (14.92%), camphor (12.50%), and terpineol (10.77%). TC-EtOH showed powerful antimicrobial activity in *C. Tropicalis* (0.03 mg/mL). The IC<sub>50</sub> values of the TC-EtOH of the DPPH were determined 21.5  $\mu$ g/mL. The IC<sub>50</sub> values were calculated 37.28 and 44.40  $\mu$ g/mL on the MG63 and MCF-7 cell lines, respectively. It was observed that the wounds treated with TC-EtOH showed a faster healing.

**Conclusions:** According to results, *T. capitata* species are thought to be natural antioxidants and a novel pharmaceutical compound for the pharmaceutical industry.

**Keywords:** antimicrobial; antioxidant; cytotoxic activity; *Thymbra capitata*; wound healing.

### Öz

**Amaç:** Bu çalışmada *Thymbra capitata* etanolik ekstraktının (TC-EtOH) kimyasal bileşimi, antimikrobiyal, antioksidan, sitotoksik aktiviteleri ve *in-vivo* yara iyileştirici etkileri araştırıldı.

**Yöntem:** TC-EtOH'nin kimyasal bileşimi gaz kromatografisi-kütle spektrometrisi (GC-MS) ile analiz edildi. Antioksidan ve antimikrobiyal özellikleri sırasıyla 2,2-difenil-1-pikrilhidrazil (DPPH), disk difüzyonu testi ve minimum inhibisyon konsantrasyon (MIC) yöntemleriyle belirlendi. Sitotoksik aktivite MG63 (insan osteosarkomu) ve MCF-7 (insan meme kansinomu) hücre hatlarında 2,3-bis-(2-metoksi-4-nitro-5-sülfopfenil)-2H-tetrazolium-5-karboxanilid (XTT) yöntemiyle belirlendi. Tümör nekroz faktörü alfa (TNF- $\alpha$ ) seviyesi ELISA metoduyla belirlendi.

\*Corresponding author: Ceylan Hepokur, Department of Biochemistry, Faculty of Pharmacy, Sivas Cumhuriyet University, 58140 Sivas, Turkey, E-mail: cozsoya@gmail.com. <https://orcid.org/0000-0001-6397-1291>

Sema Misir, Department of Biochemistry, Faculty of Pharmacy, Sivas Cumhuriyet University, 58140 Sivas, Turkey

Tutku Tunç, Department of Microbiology, Faculty of Pharmacy, Sivas Cumhuriyet University, 58140 Sivas, Turkey

Ugur Tutar, Department of Botanica, Faculty of Pharmacy, Sivas Cumhuriyet University, 58140 Sivas, Turkey

Ali Ihsan Hepokur, NETA OSGB, 58000 Sivas, Turkey

Mehmet Çiçek, Department of Biology, Pamukkale University, Faculty of Arts and Sciences, 20070 Denizli, Turkey

**Bulgular ve Tartışma:** TC-EtOH'de bulunan ana bileşenler tetratriakontan (%14,92), kafur (%12,50), terpineol (%10,77)'dür. TC-EtOH, *Candidatropicalis*'e (0,03 mg/mL) karşı güçlü bir antimikrobiyal aktivite gözlemlendi. TC-EtOH'nin DPPH için IC<sub>50</sub> değeri 21,5 µg/mL olarak belirlendi. TC-EtOH'nin MG63 ve MCF-7 hücre serileri için IC<sub>50</sub> değerleri sırasıyla 37,28 µg/mL ve 44,40 µg/mL olarak belirlendi. *In vivo* çalışmalardan elde edilen bulgulara göre, TC-EtOH ile tedavi edilen yaraların kontrol grubuna göre daha hızlı iyileşme gösterdiğini gözlemlendi.

**Sonuç:** *T. capitata* türleri doğal antioksidan ve ilaç endüstrisi için yeni bir farmasötik bileşik olabilirler.

**Anahtar kelimeler:** Antimikrobiyal; Antioksidan; Sیتو-toksik aktivite; *Thymbra capitata*; Yara iyileşmesi.

## Introduction

Aromatic plants have been used in traditional medicine since the since ancient times [1]. These plants contain essential oils, terpenes, and other non-terpene components [2]. Natural products have been used to prevent and treat the progression of many diseases [3]. Lamiaceae family is the most be aromatic species of great interest [4]. Among these species, thymus, origanum, satureja, thymbra, and coridothymus types are especially important in terms of their wide distribution and economic benefits [5]. In the worldwide, Thymbra herb is categorized into four species; (*Thymbra calostachya* (Rech.f.) Rech.f., *Thymbra capitata* (L.) Cav., *Thymbra sintenisii* Bornm. and Azn., *Thymbra spicata* L.). These species show the greatest variety in the Eastern Mediterranean region. It is widely distributed from Portugal coast (the beginning of the Mediterranean) to Iraq, from Egypt and Algeria to the Black Sea [6]. In Turkey, Thymbra species are represented into five taxon, which are *T. capitata* (L.) Cav., *T. sintenisii* Bornm and Azn. subsp. *sintenisii*, *T. sintenisii* Bornm and Azn. subsp. *isaurica* P. H. Davis, *T. spicata* L. subsp. *intricate* [5], R. Morales, *T. spicata* L. subsp. *Spicata* [7]. Thymbra species have antimicrobial, antioxidant, anti-inflammatory, antiparasitic, and antiproliferative activities, among other beneficial biological effects [8–10]. In the literature, there are many studies examining the beneficial biological effects of Thymbra species [3, 11–13]. However, to the best of our knowledge, there are few reports published about the *T. capitata* strain.

The aim of this study was to determine the phytochemical composition, antioxidant, antimicrobial, and cytotoxic activities of *T. capitata* ethanolic extract (TC-EtOH) and to investigate its probable preventive effects.

## Materials and methods

### Plant material

*T. capitata* was gathered from Denizli, province in the Southwestern Turkey, in July 2014. M. Çiçek has confirmed the taxonomic recognition of plant. A voucher specimen (*T. capitata*; Herbarium No: 2014-33) was deposited at the Çiçek Herbarium in Department of Biology, Faculty of Arts and Science, Pamukkale University, Denizli, Turkey (PAU-M. Çiçek Herb.). Collection information of *T. capitata*; C2 Denizli: 31 km from Kale to Muğla, 945 m, *Pinusbrutia* clearings, 13.07.2014, M. Çiçek 2014-33 (Herb. M. Çiçek).

### Preparation of the extract

Aerial parts of plants have parted after drying in the dark under sterile conditions. Plant samples were grinded before extraction process. Twenty grams of the sample were added in 40 mL ethanol for extraction. Ethanol was evaporated using a rotary evaporator (Stuart RE300) under reduced pressure at 30 °C [14].

### GC-MS analysis

Chemical composition of TC-EtOH was determined with the method modified by Abay [15]. Agilent Technologies GC 7890A, equipped with 5975 Triple Axis Detector mass spectrometer was employed to perform gas chromatography-mass spectrometry (GC-MS) analysis. DB-WAXetr column (60 m × 0.20 mm × 0.25 µm), electron ionization system and ionization energy of 70 eV were used for GC-MS detection. Helium was the carrier gas at a flow rate of 1 mL/min. The GC oven temperature was kept at 40 °C for 5 min and programmed to 250 °C at a rate of 5 °C/min and kept constant at 250 °C for 20 min. The library search was carried out using NIST.

### Antioxidant activity

Radical scavenging activity of extracts was performed with the method modified by Ou et al. [16]. Different concentrations of the extracts were prepared, and then equal volumes (1,000 µL) of 2,2-diphenyl-1-picrylhydrazyl (DPPH) and sample solutions were mixed and incubated for 30 min. After incubation absorbance was measured by spectrophotometer at 517 nm. The ascorbic acid as an antioxidant was used to compare the results.

### Microbiological activity

*Klebsiella pneumoniae* (ATCC 10031), *Shigellaboydii* (ATCC 9905), *Pseudomonas aeruginosa* (ATCC 27853), *Proteus vulgaris* (ATCC 7829), *Staphylococcus aureus* (ATCC 25923), *Bacillus cereus* (ATCC 10987), and *Candida tropicalis* (ATCC 750) were used to determine antimicrobial and antifungal properties of TC-EtOH. All bacteria and fungi were obtained from American Type Culture Collection (ATCC).

### Disc diffusion assay

The antimicrobial activity of TC-EtOH was evaluated by using disc diffusion method [17], in which 100  $\mu\text{L}$  of suspension containing  $10^8$  CFU/mL of bacteria and  $10^6$  CFU/mL of fungi spread on Mueller Hinton Agar (MHA) and Potato Dextrose Agar (PDA) were used, respectively. After the impregnation of the disc (6 mm diameter) with 20  $\mu\text{L}$  at 50 mg/mL concentration, it was added on the inoculated agar. Ethanol was used as negative control. Commercially available cefoperazone/sulbactam (105  $\mu\text{g}$ ) and fluconazole (25  $\mu\text{g}$ ) discs were employed for positive control of the bacteria and fungi, respectively. Regarding the zones of growth inhibition around the disks; bacteria's were measured after 24 h incubation at 37 °C, whereas fungi's were measured after 28 h incubation at 37 °C. All the assays were done in triplicate [18].

### Micro-well dilution assay

Antimicrobial activities of TC-EtOH were determined by broth micro dilution technique, which was performed according to CLSI protocol [19]. Minimal Inhibitory Concentration (MIC) values of TC-EtOH were performed by micro-well dilution method. Sample concentration range (0.03–2 mg/mL) was prepared with Müller–Hinton Broth (MHB). The suspensions were adjusted to 0.5 McFarland standard turbidity. Ninety five microliters of MHB and 5  $\mu\text{L}$  of the inoculums were added into each well; whereas 195  $\mu\text{L}$  of Nutrient broth without compound and 5  $\mu\text{L}$  of the inoculum were used as negative control. Piperacillin/tazobactam (8/1) and fluconazole were used as positive control for bacteria and fungi, respectively.

### In vivo wound healing activity

This research was carried out in Sivas Cumhuriyet University Animal Laboratory in accordance with the guidelines of Sivas Cumhuriyet University Animal Experiments Local Ethics Committee. The research was conducted in accordance with ethical rules with the decision of Sivas Cumhuriyet University Animal Experiments Local Ethics Committee dated 06.08.2015 and number 65202830–050.04.04/69.

Male Wistar albino rats ( $n=16$ ) weighed (250–275 g) were used. Standard laboratory diet and requested drinking water were used to feed all animals, so that they can always be internal. Rats were housed such that eight animals per cage. Rats capable of normal activity in cages at  $22 \pm 2$  °C, humidity (50–70%) and on 12 h day/night set will be kept in the room [20]. The anesthesia of the rats was performed with ketamine hydrochloride (90 mg/kg). Anaesthetized animals were made 3 cm full thickness incision and a punch biopsy was opened  $1 \times 1$  cm<sup>2</sup>. Two groups of eight rats were formed by randomly assigning the animals. While one was the control group, the other group was treated with TC-EtOH. Sterile solution of extracts was applied topically to each wound of the animals at a dose of 50  $\mu\text{L}$ /wound once daily for seven days. The animals of all groups were euthanized in accordance with the procedures on seventh day, their cardiac blood and tissue samples were taken. Blood samples were centrifuged at  $1500 \times g$  for 10 min. TNF- $\alpha$  values were determined by ELISA according to the manufacturer's instructions (BOSTER, Rat TNF $\alpha$  ELISA Kit, Cat No:EK0526) [21]. Tissue samples were examined histopathologically. The changes in the injured area were regularly measured and the data were calculated according to the following Eqn.: wound

contraction=(healed area/total wound area)  $\times$  100. The wound healing was compared to the control group (healed area = wound area – present wound area) [22].

### Cell culture

Human osteosarcoma (MG63, ATCC-CRL-1427), human breast adenocarcinoma (MCF-7, ATCC-HTB-22), and mouse fibroblast (ATCC-CRL-6364) cell lines were purchased from ATCC. Cells were grown at 37 °C in a humidified incubator in 5% CO<sub>2</sub>. All media were supplemented with 1% penicillin (100 U/mL) and streptomycin (100  $\mu\text{g}$ /mL), and 10% fetal bovine serum (FBS). Live cell as control was considered as 100%.

### Cytotoxicity assay

2,3-bis-(2-methoxy-4-nitro-5-sulphophenyl)-2H-tetrazolium-5-carboxanilide (XTT) method was used to evaluate cytotoxic activity. All cells were seeded in a 96-well plate in growth medium then treated with different concentrations of test compounds and incubated in a humidified CO<sub>2</sub> atmosphere at 37 °C for 24 h. Following the incubation, 100  $\mu\text{L}$  XTT solution was added to each well for another 2 h incubation. The optical density values were measured at 475 nm with a microplate reader [23].

### Statistical analysis

Data were expressed as mean  $\pm$  standard deviation [ $\pm$ SD]. Statistical analysis were performed using the Sigma Plot 12.0. All determinations were computed independent triplicated [ $n=3$ ].

## Results and discussion

Since ancient times, aromatic plant species, which are defined as “thyme” have been used to treat a range of symptoms and complaints in different diseases [24–26]. Lamiaceae species possess antimicrobial, antioxidant, and antitumoral activities [27].

### Determining chemical composition of TC-EtOH

Phytochemical composition of TC-EtOH was analyzed by GC-MS. The TC-EtOH has approximately 53 different compounds (Table 1), and approximately 90% of compounds were identified (Figure 1). Tetratria contane (14.92%), camphor (12.50%), terpineol (10.77%) were major components of TC-EtOH. Machado et al. investigated chemical composition of *T. capitata*, and they reported that the carvacrol was as the dominant compound in *T. capitata* [28]. Another study, Ali et al. demonstrated that terpinene (5.8%), *p*-cymene (7.7%), and carvacrol (66%) were the

major components in these species. Miguel et al. reported that carvacrol, thymol, *p*-cymene, thymol, and camphor were as major compounds of the *Thymbra* and *Tymus* species [8]. Barberan et al. showed that *T. capitata* species have high amount of flavonoid [29]. Our results were different from the previous reports. This situation may arise from geographical origin, extraction methods, as well as the harvest time [27].

### Determination of the antioxidant activity of TC-EtOH

DPPH method is widely used to determine for scavenging free radicals [30]. Radical scavenging activity of TC-EtOH (IC<sub>50</sub>) was found to be 21.5 µg/mL, while IC<sub>50</sub> value of the ascorbic acid was found to be 5.02 µg/mL. Galego et al. have examined on of *T. capitata*'s essential oils gathered from Portugal region [24]. The comparison of DPPH activity according to butylated hydroxyanisole (BHA) showed that it has high antioxidant scavenging activity. Saidi et al. have reported that the DPPH value of essential oil from *T. capitata* as 1.28 µmol/mL [31]. Faleiro et al. have studied antioxidant and antimicrobial properties of *T. capitata* (*L.*) *Cav.* IC<sub>50</sub> value was found to be 27.84 µg/mL [25]. It is clear that results of our radical scavenging activity were similar with previous reports.

### Determination of the antimicrobial activity of TC-EtOH

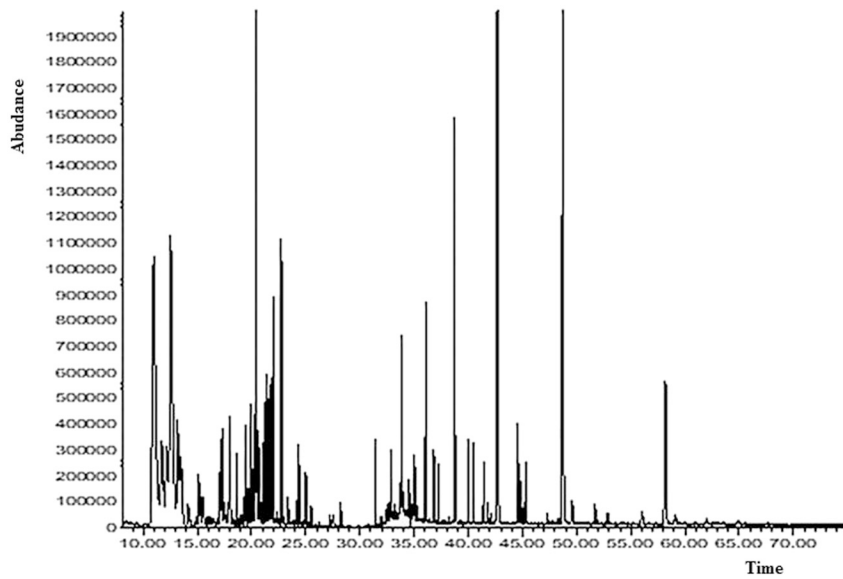
The activity was assessed by the presence or absence of inhibition zones and MIC values. The MIC values detected at the lowest concentrations without visible growth were defined as concentrations completely inhibiting microbial growth (Table 2). Results showed that TC-EtOH exhibited antibacterial activity against *C. tropicalis* (0.03 mg/mL), and also against bacterial strains (0.25–1 mg/mL). Previous studies have reported that various extracts obtained from *T. capitata* herb were effective on microorganisms [32,33].

### Determining the wound healing activity of TC-EtOH

In comparison to the control group, the treated group showed a more accelerated wound healing process in the rats [Healed area = wound area - present wound area, (Supplementary 1)]. In the process of wound healing, reformation of vascular structures is very important for

**Table 1:** GC-MS identified phytochemicals present in *Thymbra capitata* ethanolic extract.

Number	RT, min	Area%	Name
1	10.96	12.52	Camphor
2	11.69	3.24	Borneol
3	12.15	2.40	Verbenyl, ethyl ether
4	12.53	10.77	α-Terpineol
5	13.18	2.80	Verbenone
6	13.47	1.96	2-Hydroxycineole acetate
7	14.15	0.64	Carvone
8	15.13	0.82	Bornyl acetate
9	15.47	0.55	Thymol
10	17.09	0.73	2-Carene-4-ol
11	17.26	0.96	α-Bourbonene
12	17.48	0.36	Methyleugenol
13	17.94	1.42	8-Hydroxycarvotanacetone
14	18.59	0.47	Aromandendrene
15	19.41	0.90	α-Bourbonene
16	19.69	0.25	Isoshyobunone
17	19.97	0.69	Caryophyllene oxide
18	20.08	0.34	α-Bourbonene
19	20.18	0.27	3-Carene, 4-isopropenyl-
20	20.31	0.56	Limonen-6-ol, pivalate
21	20.42	9.00	Caryophyllene oxide
22	20.54	0.56	α-Gurjunene
23	20.70	1.02	Viridiflorol
24	20.99	0.15	Santalol
25	21.16	0.96	Tetracyclo[6.3.2.0(2,5).0(1,8)]tridecan-9-ol, 4,4-dimethyl-
26	21.44	1.17	Longipinocarveol
27	21.63	0.81	Caryophyllene oxide
28	21.72	0.14	Dodecane, 5,8-diethyl-
29	21.89	0.84	5,6,6-Trimethyl-5-(3-oxobut-1-enyl)-1-oxaspiro[2.5]octan-4-one
30	22.05	1.46	Isoaromadendrene epoxide
31	22.74	1.91	Acetic acid, 2,6,6-trimethyl-3-methylene-7-(3-oxobutylidene)oxepan-2-yl ester
32	23.39	0.26	Heptadecane
33	24.09	0.18	α-Murolene
34	24.35	0.71	Hexahydrofarnesyl acetone
35	25.51	0.11	Octadecane
36	28.21	0.24	Hexadecanoic acid, ethyl ester
37	31.39	0.45	Heneicosane
38	32.48	0.08	Ethyl Oleate
39	32.77	0.08	Stearic acid, ethyl ester
40	32.83	0.30	Heptadecane, 9-hexyl-
41	33.13	0.06	13-Heptadecyn-1-ol
42	33.79	0.24	1-Eicosanol
43	33.90	0.73	Pentacosane
44	34.55	0.26	4,8,12,16-Tetramethylheptadecan-4-olide
45	36.05	1.05	Hexacosane
46	37.28	0.40	Heptacosane
47	38.74	2.76	Octacosane
48	40.00	0.63	Nonacosane
49	40.48	0.60	Hentriacontane
50	41.78	0.17	Hentriacontane
51	42.68	14.92	Tetratriacontane
52	51.69	0.31	Tetracontane
53	58.19	3.09	Tetratetracontane



**Figure 1:** The major components identified by gas chromatography-mass spectrometry from samples of TC-EtOH. Chromatographic profiles of samples from *Thymbra capitata*.

**Table 2:** Antimicrobial screening of TC-EtOH. Antimicrobial activity of TC-EtOH performed by disc diffusion and micro-well dilution assay. Cefoperazone/sulbactam and fluconazole were used for positive control of the bacteria and fungi, respectively.

Bacteria	Zone of Inhibition, mm		MIC, mg/mL	
	Extract	Control <sup>a</sup>	Extract	Control <sup>b</sup>
<i>Klebsiella pneumoniae</i>	22.3 ± 0.5	26.6 ± 2.0	0.5	<0.03
<i>Shigella boydii</i>	17.3 ± 0.5	25.0 ± 1.7	0.5	<0.03
<i>Pseudomonas aeruginosa</i>	11.3 ± 1.5	24.6 ± 1.1	1.0	<0.03
<i>Proteus vulgaris</i>	14.0 ± 1.0	28.6 ± 1.1	1.0	<0.03
<i>Staphylococcus aureus</i>	29.6 ± 2.0	19.6 ± 1.5	0.25	<0.03
<i>Bacillus cereus</i>	16.0 ± 1.7	32.3 ± 0.5	0.5	<0.03
Fungus	Extract	Control <sup>c</sup>	Extract	Control <sup>d</sup>
<i>Candida tropicalis</i>	12.8 ± 0.4	13.1 ± 0.8	0.03	<0.03

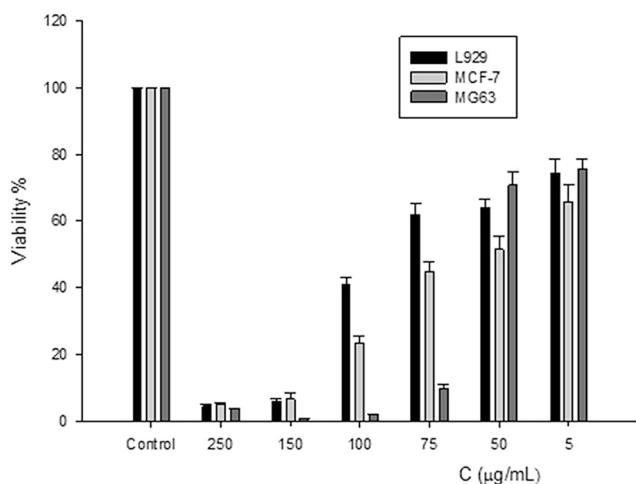
<sup>a</sup>Cefoperazone/ Sulbactam 2:1, <sup>b</sup> Piperasilin/Tazobactam(8:1), <sup>c</sup>Fluconazol disk, <sup>d</sup>FluconazolMIC, minimal inhibitory concentration.

the nutrition of the damaged tissue. Angiogenesis is to be closely associated with inflammation. Monocyte/macrophages are essential for the wound healing and the accumulation of monocyte/macrophage in the region of the wound is very important for capillary organization and collateral formation. These inflammatory cells mediate the formation of a series of angiogenic cytokine and growth factors. Therefore, determination of TNF- $\alpha$  level would be an important determinant for the wound healing [34]. In this study, using an animal model of excisional wound healing, we found that TC-EtOH application caused significant healing in all wound models compared to the control group (Supplementary 2). The capability TC-EtOH is crucially dependent on the modulation of the inflammatory factor TNF- $\alpha$ . The changes in the levels of TNF- $\alpha$  was examined and we found that the level of TNF- $\alpha$  levels decreased significantly.

Mohamad et al. showed that the ethanolic extract of *T. spicata* has wound healing activity [35]. Tutar et al. demonstrated that the ethanol extract of *T. sintonisii* was reduced the total wound area [36].

### Determination of the cytotoxic activity of TC-EtOH

Cancer is a pathological condition that occurs with a genetic and developmental process [37]. Anticancer drugs have toxicity and serious side effects for normal cells due to the chronic use [38]. Developing a new generation of anticancer agents became a quite popular research area. In the study, the cytotoxic activity of TC-EtOH was tested on MCF-7, MG63, and L929 cells. The cell proliferation



**Figure 2:** Cell cytotoxicity assessment by XTT assay. The data represents the viability of MCF-7, MG63, and L929 cells treated for 24 h with TC-EtOH. The experiments were performed at three times for each experimental group. Data are shown as the mean  $\pm$  SD.

percentage of the control group has been considered as 100%.  $IC_{50}$  values were found of TC-EtOH 37.28, 44.40 and 44.84  $\mu\text{g/mL}$  on MG63, MCF-7, and L929 cells, respectively (Figure 2). TC-EtOH was reduced cell viability on MG63 and MCF-7 cells a dose-dependent manner. It has been observed that the TC-EtOH was more effective on MG63 than MCF-7 cells. Miguel et al. showed that *T. capitata* has antiproliferative activity on THP-1 cells for 24 h. The essential oils from *T. capitata* were decreased viability of THP-1 cells in a dose-dependent manner [8]. Delgado-Adamez et al. investigated the cytotoxic activities of essential oil of *T. capitata* and Thymus species on HeLa (adherent cells) and U937 (free-floating cells). They reported that these essential oils were decreased cell viability of both tumor cells in a dose-dependent [33]. Alexa et al. reported that Thyme and sage essential oils exhibited antiproliferative activity on A375 human melanoma and B164A5 mouse melanoma cells [27]. The bioactive compounds of thymus endemic species show antiproliferative activities against on two human lung cancer cell lines (A549 and NCI-H226) [39], human colon cancer (HCT116) cells [40], A375 human melanoma cells [41], MCF-7-breast and LNCaP-prostate cancer cell lines [42].

## Conclusion

The results of this study revealed that the TC-EtOH established interesting biological effects. TC-EtOH displayed strong antioxidant and antimicrobial activities. Moreover, results clearly showed that the TC-EtOH has *in vitro*

cytotoxic activity and *in vivo* wound healing activity. However, more studies are needed to elucidate their mechanism in this area.

**Acknowledgments:** None.

**Research funding:** None declared.

**Author contributions:** All authors have accepted responsibility for the entire content of this manuscript and approved its submission.

**Competing interests:** Authors state no conflict of interest.

## References

1. Edris AE. Pharmaceutical and therapeutic potentials of essential oil sand their individual volatile constituents: a review. *Phytother Res* 2007;21:308–23.
2. Bakkali F, Averbeck S, Averbeck D, Idaomar M. Biological effects of essential oils-a review. *Food Chem Toxicol* 2008;46:446–7.
3. Fitsiou E, Anastopoulos I, Chlichlia K, Galanis A, Kourkoutas I, Panayiotidis MI, et al. Antioxidant and antiproliferative properties of the essential oils of *Satureja thymbra* and *Satureja parnassica* and their major constituents. *Anticancer Res* 2016;36: 5757–63.
4. Saija AA, Speciale D, Trombetta CL, Tuttolomondo T, Bella SL, Licata M, et al. Phytochemical, ecological and antioxidant evaluation of wild Sicilian thyme: *Thymbra capitata* (L.) Cav. *Chem Biodivers* 2016;13:1641–55.
5. Davis PH *Flora of Turkey and the East Aegean Islands*, vol. 7. Edinburgh: Edinburgh University Press; 1982:382–4 pp.
6. Erken S. Morphological and anatomical studies on *Thymbra sintenisii* Bornm. *And Azn. Labiatae*. *Turk J Bot* 2005;29:389–97.
7. Jalas J. *Flora of Turkey and the East Aegean Islands*, vol. 7. Edinburgh: Edinburgh University Press; 1982:382–4 pp.
8. Miguel M, Gago C, Antunes M, Megías C, Giraldo I, Vioque J. Antioxidant and antiproliferative activities of the essential oils from *Thymbra capitata* and Thymus species grown in Portugal. *Evid Based Complement Alternat Med* 2015;2015:851721.
9. Cardile V, Russo A, Formisano C, Rigano D, Senatore F, Arnold NA, et al. Essential oils of *Salvia bracteata* and *Salvia rubifolia* from Lebanon: chemical composition, antimicrobial activity and inhibitory effect on human melanoma cells. *J Ethnopharmacol* 2009;126:265–72.
10. Russo A, Formisano C, Rigano D, Cardile V, Arnold NA, Senatore F. Comparative phytochemical profile and antiproliferative activity on human melanoma cells of essential oils of three Lebanese *Salvia* species. *Ind Crop Prod* 2016;83:492–99.
11. Dandlen SA, Lima AS, Mendes MD, Miguel MG, Faleiro ML, Sousa MJ. Antioxidant activity of six Portuguese thyme species essential oils. *Flavour* 2010;25:150–55.
12. Figueiredo AC, Barroso JG, Pedro LG, Salgueiro L, Miguel MG, Faleiro ML. Portuguese *Thymbra* and *Thymus* species volatiles: chemical composition and biological activities. *Curr Pharmaceut Des* 2008;14:3120–40.
13. Kiliç T. Analysis of essential oil composition of *Thymbra spicata* var. *spicata*: antifungal, antibacterial and antimycobacterial activities. *Z Naturforsch C Biosci* 2006;61:324–8.

14. Alzoreky NS, Nakahara K. Antibacterial activity of extracts from some edible plants commonly consumed in Asia. *Int J Food Microbiol* 2003;80:223–30.
15. Abay G, Altun M, Karakoç UC, Gül F, Demirtas I. Insecticidal activity of fatty acid-rich Turkish bryophyte extracts against *Sitophilus granarius* (Coleoptera: Curculionidae). *Comb Chem High Throughput Screen* 2013;16:806–16.
16. Ou B, Huang D, Hampsch-Woodill M, Flanagan JA, Deemer EK. Analysis of antioxidant activities of common vegetables employing oxygen radical absorbance capacity (ORAC) and ferric reducing antioxidant power (FRAP) assays: a comparative study. *J Agric Food Chem* 2002;50:3122–8.
17. Wayne PA. Clinical Laboratory Standards Institute. Performance standards for antimicrobial disk susceptibility tests. Approved standard, 9th ed. Clinical Laboratory Standards Institute: CLSI document; 2006. M2-A9. 26:1.
18. Selim AS, Adam ME, Hassan SM, Albalawi AR. Chemical composition, antimicrobial and antibiofilm activity of the essential oil and methanol extract of the Mediterranean cypress (*Cupressus sempervirens* L.). *BMC Compl Alternative Med* 2014; 14:179.
19. Wayne PA, Clinical and Laboratory Standards Institute (CLSI). Performance standards for antimicrobial susceptibility testing. Twenty-Fourth Informational Supplement. CLSI Document; 2014. M100-S24.
20. Gopinath D, Rafiuddin AM, Gomathi K, Chitra K, Sehgal PK, Jayakumar P. Dermal wound healing processes with curcumin incorporated collagen films. *Biomaterials* 2004;25:1911–7.
21. Wen X, Han XR, Wang YJ, Fan SH, Zhang ZF, Wu DM, et al. Effects of S100A12 gene silencing on serum levels of anti-inflammatory/pro-inflammatory cytokines in septic rats through the ERK signaling pathway. *J Cell Biochem* 2018;119: 4038–49.
22. Murthy S, Gautam MK, Shalini G, Purohit V, Sharma H, Goel RK. Evaluation of *in vivo* wound healing activity of *Bacopa monniera* on different wound model in rats. *BioMed Res Int* 2013;29: 972028.
23. Loizzo MR, Tundis R, Menichini F, Saab A, Statti G, Menichini F. Cytotoxic activity of essential oils from Labiatae and Lauraceae families against *in vitro* human tumor models. *Anticancer Res* 2007;27:3293–9.
24. Galego L, Almeida V, Gonçalves V, Costa M, Monteiro I, Matos F, et al. Antioxidant activity of the essential oils of *Thymbra capitata*, *Origanum vulgare*, *Thymus mastichina* and *Calamintha baetica*. *Acta Hort* 2008;765:325–34.
25. Faleiro L, Miguel G, Gomes S, Costa L, Venâncio F, Teixeira A. Antibacterial and antioxidant activities of essential oils isolated from *Thymbra capitata* L. (Cav.) and *Origanum vulgare* L. *Agric Food Chem* 2005;53:8162–68.
26. Ravid U, Putievsky E. Composition of essential oils of *Thymbra spicata* and *Satureja thymbra* Chemotypes 1. *Planta Med* 1985;51: 337–8.
27. Alexa E, Sumalan RM, Danciu C, Obistoiu D, Negrea M, Poiana MA, et al. Synergistic antifungal, allelopathic and anti-proliferative potential of *Salvia officinalis* L., and *Thymus vulgaris* L. essential oils. *Molecules* 2018;16:23.
28. Machado M, Dinis AM, Salgueiro L, Cavaleiro C, Custódio JB, Sousa Mdo C. Anti-Giardia activity of phenolic-rich essential oils: effects of *Thymbra capitata*, *Origanum virens*, *Thymus zygis* subsp. *sylvestris*, and *Lippia graveolens* on trophozoites growth, viability, adherence, and ultrastructure. *Parasitol Res* 2010;106: 1205–15.
29. Barberán FAT, Hernández L, Tomás F. A chemotaxonomic study of flavonoids in *Thymbra capitata*. *Phytochemistry* 1986;25:561–2.
30. Kedare SB, Singh RP. Genesis and development of DPPH method of antioxidant assay. *J Food Sci Technol* 2011;48:412–22.
31. Saidi M, Ghafourian S, Zarin-Abadi M, Movahedi K, Sadeghifard N. *In vitro* antimicrobial and antioxidant activity of black thyme (*Thymbra spicata* L.) essential oils. *Rom Arch Microbiol Immunol* 2012;71:61–9.
32. Palmeira-de-Oliveira A, Gaspar C, Palmeira-de-Oliveira R, Silva-Dias A, Salgueiro L, Cavaleiro C, et al. The anti-candida activity of *Thymbra capitata* essential oil: effect upon pre-formed biofilm. *J Ethnopharmacol* 2012;140:379–83.
33. Delgado-Adámez M, Garrido M, Bote ME, Fuentes-Pérez MC, Espino JD. Chemical composition and bioactivity of essential oils from flower and fruit of *Thymbra capitata* and *Thymus* species. *J Food Sci Technol* 2017;54:1857–65. Available from: [https://www.ncbi.nlm.nih.gov/pubmed/?term=Mart%C3%ADn-Vertedor%20D%5BAuthor%5D&cauthor=true&cauthor\\_uid=28720941Martín-Vertedor](https://www.ncbi.nlm.nih.gov/pubmed/?term=Mart%C3%ADn-Vertedor%20D%5BAuthor%5D&cauthor=true&cauthor_uid=28720941Martín-Vertedor).
34. Schultz GS, Chin G, Moldawer L, Diegelmann RF. Principles of wound healing, mechanisms of vascular disease: a reference book for vascular specialists. Robert Fitzridge and Matthew Thompson edition; 2011. Available from: [https://www.ncbi.nlm.nih.gov/books/NBK534260/pdf/Bookshelf\\_NBK534260.pdf](https://www.ncbi.nlm.nih.gov/books/NBK534260/pdf/Bookshelf_NBK534260.pdf).
35. Khalil M, Khalifeh H, Baldini F, Salis A, Damonte G, Daher A, et al. Antisteatotic and antioxidant activities of *Thymbra spicata* L. extracts in hepatic and endothelial cells as *in vitro* models of non-alcoholic fatty liver disease. *J Ethnopharmacol* 2019;15:111919.
36. Tutar U, Hepokur C, Misir S, Hepokur AI, Duman F. Antimicrobial, antioxidant, cytotoxic and wound healing effects of *Thymbra sintonensis* extract. *Indian J Pharmaceut Sci* 2018;80:868–74.
37. Kuno T, Tsukamoto T, Hara A, Tanaka T. Cancer chemoprevention through the induction of apoptosis by natural compounds. *JBPC* 2012;3:156–73.
38. Tan W, Lu J, Huang M, Li Y, Chen M, Wu G, et al. Anticancer natural products isolated from Chinese medicinal herbs. *Chin Med* 2011;6:27.
39. Privitera G, Napoli E, Luca T, Ruberto G, Castorina S. *In vitro* anti-proliferative effect of *Salvia officinalis* essential oil and its three main components on human lung cancer cells. *Am J Phytomed Clin Ther* 2014;2:1159–68.
40. Itani WS, El-Banna SH, Hassan SB, Larsson RL, Bazarbachi A, Gali-Muhtasib HU. Anti colon cancer components from lebanese sage (*Salvia libanotica*) essential oil: mechanistic basis. *Canc Biol Ther* 2008;7:1765–73.
41. Bendif H, Boudjeniba M, Miara MD, Biqiku L, Bramucci M, Lupidi G, et al. Essential oil of *Thymus munbyanus* subsp. *coloratus* from Algeria: chemotypification and *in vitro* biological activities. *Chem Biodivers* 2017;14: e1600299.
42. Hussain AI, Anwar F, Chatha SAS, Latif S, Sherazi STH, Ahmad A, et al. Chemical composition and bioactivity studies of the essential oils from two *Thymus* species from the Pakistani flora. *LWT – Food Sci Technol (Lebensmittel-Wissenschaft-Technol)* 2013;50:185–92.

**Supplementary Material:** The online version of this article offers supplementary material (<https://doi.org/10.1515/tjb-2019-0470>).