#### RESEARCH



# Comparison of hyaluronic acid, hypochlorous acid, and flurbiprofen on postoperative morbidity in palatal donor area: a randomized controlled clinical trial

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Received: 1 July 2022 / Accepted: 26 December 2022 / Published online: 3 January 2023 © The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2023

#### Abstract

**Objective** This study aims to evaluate the effects of topical hyaluronic acid (HA), hypochlorous acid (HOCl), and flurbiprofen on postoperative morbidity of palatal donor sites after free gingival graft (FGG) surgery.

**Materials and methods** Sixty patients requiring FGG were randomly assigned into four groups: control, HA gel (600 mg/100 g high molecular weight hyaluronic acid), HOCl spray (170–200 ppm, ph7.1), flurbiprofen spray (0.075gr flurbiprofen). Topical agents were applied for 14 days, according to groups. Patients were followed for 28 days. Palatal healing was assessed with the Laundry wound healing index (WHI). Complete epithelization (CE) was evaluated with photographs and  $H_2O_2$  bubbling. Pain, burning sensation, chewing efficacy, and tissue color match (CM) were evaluated using a visual analog scale (VAS). Postoperative analgesic consumption and delayed bleeding (DB) were also recorded.

**Results** HA provided better WHI values on the 7<sup>th</sup>, 14<sup>th</sup>, and 21<sup>st</sup> days compared to the other groups, respectively (p < 0.05). CE was formed on the 21<sup>st</sup> day in the HA group but on the 28<sup>th</sup> day in the other groups. HOCl and flurbiprofen groups were not different from the control group or each other in terms of WHI. HOCl had the lowest VAS scores of all time periods. DB was not observed in any group. Significantly fewer analgesics were taken in the topical agent-applied groups compared to the control group.

**Conclusions** HA exhibits a positive impact on the epithelization of palatal wound healing and color matching. HOCl and flurbiprofen provided less pain; however, they might have negative effects on palatal wound healing.

**Clinical relevance** As a result of obtaining free gingival grafts from palatal tissue for mucogingival surgical procedures, secondary wound healing of the donor area occurs. This wound in the palatal region can cause discomfort and pain every time patients use their mouths. The use of HA can reduce postoperative complications by accelerating wound healing and reducing pain. The topical use of flurbiprofen and HOCl can reduce patients' pain.

Keywords  $Operative \cdot Pain \cdot Plastic periodontal surgery \cdot Postoperative complications \cdot Surgical procedures \cdot Wound healing$ 

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# Introduction

Free grafts from the palate have been used most commonly for gingival augmentation. It has been stated in the literature that free gingival graft (FGG) is one of the most reliable and frequently used methods to increase the amount of keratinized gingiva. In addition, this technique prevents and treats the progression of gingival recession, can eliminate aesthetic problems, and increases vestibular depth [1]. FGG leaves an open wound site due to the removal of the epithelial layer in the palatal region and heals in two to four weeks with secondary wound healing [2, 3]. However, some complications may occur at the recipient site. Excessive bleeding, postoperative bone exposure, bone exostoses, case of mucocele, severe postoperative pain, and recurrent herpetic lesions are the most documented complications [4–6]. Even if various materials with mechanical protection [7], platelet-rich fibrin (PRF) [8], herbal extract [5], and chemotherapeutic agents [9] are used in the donor area to prevent postoperative complications, there is no definite consensus.

Flurbiprofen, 2(3-fluoro-4-phenyl–phenyl)-propionic acid, is a non-steroidal anti-inflammatory drug (NSAID) that has been shown to be safe and effective in relieving postoperative pain [10, 11]. Flurbiprofen in the form of an oral spray can be found commercially and simply applied to the palatal region directly. It has been found to inhibit the increase of pro-inflammatory cytokines such as tumor necrosis factor (TNF)- $\alpha$ , and interleukin (IL)-1 $\beta$  [12, 13], also reducing IL-6 expression [14] and inhibiting cyclooxygenase-2 (COX-2) expression [15].

Hypochlorous acid (HOCl) exhibits potent antimicrobial activities against a wide variety of microorganisms [16, 17]. HOCl, a naturally occurring molecule produced by neutrophils to destroy pathogens [18], is used in cosmetic and medical dermatological procedures due to its lack of microbe resistance, safety, and antimicrobial activity [17].

Hyaluronic acid (HA) is a natural polymer of glycosaminoglycans (GAGs) found in the joint synovial fluid and the extracellular matrix of the skin [19, 20]. It is naturally secreted during the proliferative phase of wound healing to stimulate migration and mitosis of fibroblasts and epithelial cells and has been shown to reduce levels of inflammatory mediators [21, 22]. It has been reported that HA has a positive effect on the healing of chronic wound ulcers of various etiologies, burns, and epithelial surgical wounds, regardless of the form in which it is administered topically (i.e., pad, cream, substrate) [23]. As a therapeutic agent, HA is used in tissue reconstruction [24], to accelerate wound healing [25, 26], in degenerative/inflammatory joint diseases, and in synovial fluid replacement [27, 28].

To the best of our knowledge, no studies have examined the effects of HOCl use on palatal wound healing after FGG surgery. The aim of this study was to compare the effects of locally applied HOCl, flurbiprofen, and HA on postoperative patient discomfort and wound healing of the palatal donor area in terms of pain, burning sensation, epithelialization, and color match after FGG surgery.

## **Materials and methods**

#### Study pattern

This study was designed as a prospective randomized controlled clinical trial with parallel groups. The population of the study consists of all patients who were referred to FGG for gingival augmentation between March 5, 2019, and March 15, 2022, at Pamukkale University Faculty of Dentistry Department of Periodontology. Procedures were explained to the participants, and all of them signed consent forms before participating in the study. The study was approved by Pamukkale University Ethical Committee (05.03.2019/05) and submitted to ClinicalTrials.gov (NCT05386667). All procedures performed in this study comply with the ethical standards of the institutional and/ or national research committees and the Declaration of Helsinki and its subsequent amendments or comparable ethical standards.

#### Inclusion criteria

- Patients with systematic conditions are classified as ASA Class I [29].
- Patients > 18 years requiring FGG surgery with ≤ 1 mm width of attached gingiva in the mandibular anterior and premolar region
- Exclusion Criteria
- Pregnancy, lactation, and taking contraceptive pills
- Oversensitivity or anaphylactic reactions that contraindicate the intervention
- Orofacial neurological symptoms
- Infections at operation zone
- Psychotropic medicine, sedative, or NSAI use can alter the sense of pain
- Pathological mental conditions (dementia, psychosis) and lack of cooperation
- Patients who refuse to sign consent forms
- Excessive gag reflex

Participants were randomly assigned to one of the 4 groups following a simple software-generated random number procedure via the "List Randomizer" application (https://www.random.org.lists). The power analysis of the study was carried out using the G\*Power software program (G\*Power v.3.1.9.2, Heinrich Heine University, Dusseldorf, Germany) accepting wound healing as the primary outcome variable. According to the power analysis of a previous study, the number of patients calculated for each group was 6, with  $\alpha = 0.05$ ,  $1 - \beta = 0.95$ , and f = 2.386 [7]. However, 15 patients for each group were included in the study, taking into account the loss of follow-ups (Fig. 1). All the patients received the initial periodontal therapy including scaling and root planning (SRP). Oral hygiene motivation was given to the patients, and they were recalled one month later. Full-mouth plaque score (FMPS) and full-mouth percentage bleeding score (FMBS) were recorded by assigning a binary score to each surface (1 for present, 0 for absent) and calculating the percentage of total tooth surfaces that

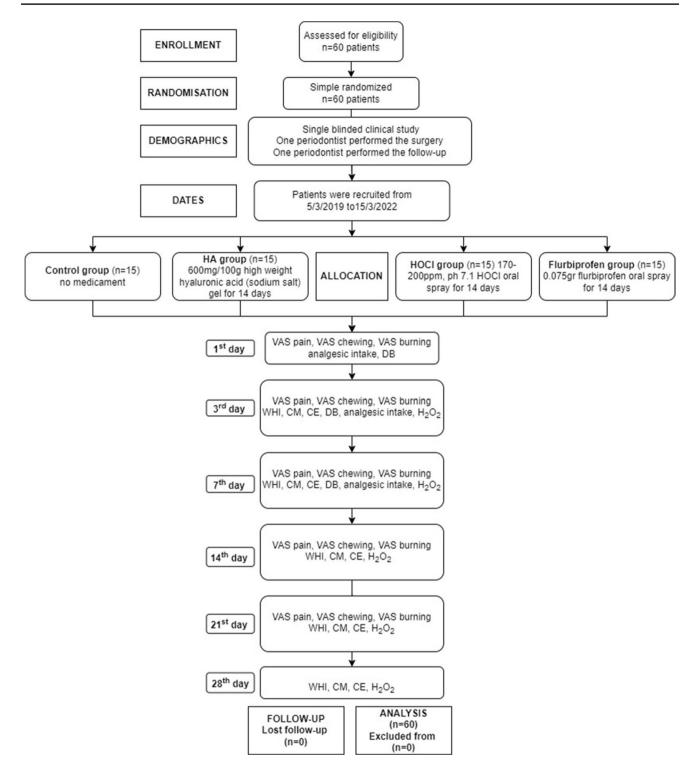


Fig. 1 Flow diagram generated in accordance with CONSORT 2010 guidelines

revealed the presence of plaque/bleeding detected by the use of a periodontal probe (Hu-Friedy, Chicago, IL, USA) [30]. The number of teeth of the patients was also recorded.

#### Study groups

 Control group: No topical agent was applied to the palatal area where the FGG was harvested.

- Hyaluronic acid (HA) group: 600 mg/100 g high molecular weight hyaluronic acid (sodium salt) gel was applied twice daily for 14 days to the palatal area where the FGG was harvested.
- Hypochlorous acid (HOCl) group: 170–200 ppm, ph. 7.1 HOCl oral spray was applied twice daily for 14 days to the palatal area where the FGG was harvested.
- Flurbiprofen group: 0.075gr flurbiprofen and additives (sorbitol, saccharin sodium, glycerin, polyoxyl 40 hydrogenated castor oil, methyl paraben, propyl paraben, ethyl alcohol, patent blue E131 (blue), menthol, sodium hydroxide) included, the oral spray was applied twice daily for 14 days to the palatal area where the FGG was harvested.

# **Surgical intervention**

In order to minimize the differences in surgical technique, all surgical procedures were performed with the same surgical technique by a single periodontist (ALA). The surgical application was applied by following the steps below briefly. After the palatal region was numbed with local infiltration anesthesia (2% lidocaine with 1:100,000 epinephrine), the area between the distal line angle of the canine and the mesial line angle of the first molar was marked as > 2 mmaway from the gingival margin, and a half-thickness incision was made. A FGG with a thickness of  $\approx 1-1.5$  mm and dimensions of  $10 \times 5$  mm was obtained from the marked area with a scalpel no: 15C. The thickness was measured in the middle of the graft with an endodontic reamer and a caliper over a flat surface. The fat and glandular tissue were removed from the graft, and then the graft was shaped to adapt to the recipient site.

## **Postoperative care**

The patients were prescribed an analgesic drug (500 mg of paracetamol). The patients in the study groups other than controls were given HA gel, HOCl, and flurbiprofen oral spray to be applied to the palatal wound areas. Patients who were received HOCl and flurbiprofen oral spray instructed to take one dose (three sprays) to the donor site twice a day for 14 days. HA gel was prescribed twice a day for 14 days to apply with its special applicator to the palatal wound area. In order not to disturb the stabilization of the clot formed in the operation area, all locally applied products were started 6-8 h after the operation. Patients were given instructions not to eat, drink, or rinse for about 30 min after spray/ gel application and advised to report the outcome if any adverse events occurred. Patients were instructed to avoid any hard brushing or trauma to the surgical site for 3 weeks. All patients were monitored, and measurements were taken by one periodontist (GTC) on days 1, 3, 7, 14, 21, and 28.

Patients were asked if they needed to use analgesics and to note the amount they used in the 7-day postoperative period.

### **Evaluated parameters**

The primary outcome of the study was to measure the palatal wound healing status using the Landry Wound Healing Index (WHI) [31], which grades the wound healing on a scale of the  $3^{rd}$ ,  $7^{th}$ ,  $14^{th}$ ,  $21^{st}$ , and  $28^{th}$  days postoperatively. This index, which has a score range of 1 (very poor) to 5 (excellent), evaluates tissue color, response to palpation, presence of granulation tissue, epithelialization of incision margins, and amount of suppuration.

The secondary outcomes of the study were to measure patients' perception of pain, discomfort while chewing, and burning sensation after FGG harvesting using a numerical rating scale (VAS) from 0 (no pain, no discomfort while chewing, no burning sense) to 10 (the worst pain imaginable, extreme discomfort while chewing, extreme burning sense) [32] on the 1<sup>st</sup>, 3<sup>rd</sup>, 7<sup>th</sup>, 14<sup>th</sup>, and 21<sup>st</sup> days, and the amount of analgesic consumption on the 1<sup>st</sup>, 3<sup>rd</sup>, and 7<sup>th</sup> days postoperatively. Tissue color match (CM) was assessed with adjacent and contralateral palatal tissue (0—no color matching to 10—excellent color matching) [32].

The patients were asked to keep a record of whether there was any bleeding in the palatal region, and at the same time, the presence of postoperative bleeding, also known as delayed bleeding (DB), was recorded as present (+) or absent (-) on the days when the patients came to the followup appointments.

The epithelialization of the palatal region was recorded according to the application of 3% hydrogen peroxide to the region and whether there was foaming or not, and was calculated as a percentage ( $H_2O_2$  bubbling) [33]. Complete epithelization (CE) was also evaluated clinically by monitoring the surface characteristics and clarity of the wound contour and recorded as "yes" or "no" [26].

## **Statistical analysis**

All data were evaluated with SPSS 21.0 (SPSS Inc., Chicago, IL). Shapiro–Wilk test was used to test the data's normality. Since the data distributions were not normal, non-parametric tests were used to evaluate the analysis. The Chi-square test was used for demographic data (age, gender), CE, and DB. The number of teeth of the participants in all groups were compared with using one-way ANOVA post hoc Tukey test. Kruskal–Wallis post-hoc Mann–Whitney U with Bonferroni correction was applied to analyze the data of different groups (FMPS, FMBS, VAS pain, chewing, burning, WHI, CM, H<sub>2</sub>O<sub>2</sub> bubbling, and analgesic consumption). The Friedman test was applied to evaluate the repeated measures within the groups. Data were expressed as min-max (median), mean ± standard deviation (SD), and frequency. p < 0.05 was considered statistically significant.

## Results

A total of 60 patients took part in the study. The long duration of this study was due to the precautions taken during the COVID-19 pandemic. There was no drop-out among the 60 patients. All patients included in the study were followed for 28 days. Demographic data for the groups are presented in Table 1. FMPS, FMBS, and the number of teeth of all patients included in the study were similar between groups (p > 0.05). The patients participating in the study did not differ in terms of age and gender (p > 0.05). None of the patients developed adverse reactions to the agents used.

When our primary outcome, WHI, was examined, the HA group was found to be better in terms of wound healing scores compared to the HOCl group on the 3<sup>rd</sup> day (p=0.019). The use of HA showed statistically better results in terms of wound healing compared to the control, flurbiprofen, and, HOCl groups on the 7<sup>th</sup>, 14<sup>th</sup>, and 21<sup>st</sup> days, respectively (Table 2, Fig. 2).

When the VAS data, which evaluated the postoperative pain levels of the patients, were analyzed, the pain levels of the patients in the HOCl group were found to be significantly lower than in the flurbiprofen group (p=0.007)and the control groups (p = 0.033) on the 1<sup>st</sup> day. On the 3<sup>rd</sup> day, patients in the HOCl group felt significantly less pain than those in the flurbiprofen (p=0.030), and control groups (p = 0.001) respectively. Also, HA and flurbiprofen administration decreased the pain levels comparing the control group (p < 0.05). All study groups showed a statistical decrease in VAS values on the 7<sup>th</sup> day compared to the control group, respectively (p < 0.05). There was no difference between any group in terms of pain on the 14<sup>th</sup> and 21<sup>st</sup>

| Table 1         Demographic data and periodontal parameters.   |                        | Control                              | HA                                   | HOCl                                 | Flurbiprofen                         | p value        |
|--|------------------------|--------------------------------------|--------------------------------------|--------------------------------------|--------------------------------------|----------------|
| full-mouth plaque score; <i>FMBS</i> ,<br>full-mouth percentage bleeding<br>score Data were presented<br>as mean $\pm$ SD, percentage.<br>p < 0.05; the significant<br>difference between groups | Women<br>Man           | 9(60%)<br>4(40%)                     | 9(60%)<br>6(40%)                     | 8(53.3%)<br>7(46.7%)                 | 8(53.3%)<br>7(46.7%)                 | 0.965          |
|  | Age<br>Number of teeth | $37.20 \pm 8.49$<br>$26.80 \pm 1.47$ | $36.93 \pm 9.04$<br>$26.60 \pm 1.72$ | $36.33 \pm 8.83$<br>$25.80 \pm 1.78$ | $38.73 \pm 9.76$<br>$26.13 \pm 1.45$ | 0.886<br>0.336 |
|  | FMPS (%)<br>FMBS (%)   | $13.30 \pm 2.21$<br>$8.63 \pm 1.47$  | $13.09 \pm 2.15$<br>$8.65 \pm 1.38$  | $12.99 \pm 1.92$<br>$8.83 \pm 1.56$  | $12.55 \pm 2.04$<br>$8.68 \pm 1.17$  | 0.928<br>0.992 |

**Table 2** WHI, CM, H<sub>2</sub>O<sub>2</sub> bubbling, and analgesic consumption of the groups. Data were presented as mean  $\pm$  SD. p < 0.05; the significant difference between groups with the different superscripts

| Parameters                             | Days                 | Control $n = 15$       | $ HA \\ n = 15 $      | HOCI $n = 15$                                    | Flurbiprofen<br>n = 15  | p value |
|--|----------------------|------------------------|-----------------------|--|-------------------------|---------|
| WHI                                    | 3 <sup>rd</sup> day  | $1.87 \pm 0.32^{a,b}$  | $2.26 \pm 0.45^{a}$   | $1.80 \pm 0.41^{b}$                              | $1.86 \pm 0.35^{a,b}$   | 0.012   |
|  | $7^{th} day$         | $2.66 \pm 0.48^{a}$    | $3.26 \pm 0.45^{b}$   | $2.46 \pm 0.51^{a}$                              | $2.53 \pm 0.51^{a}$     | 0.001   |
|  | 14 <sup>th</sup> day | $3.73 \pm 0.45^a$      | $4.26\pm0.45^{\rm b}$ | $3.46 \pm 0.51^{a}$                              | $3.47 \pm 0.52^{a}$     | 0.001   |
|  | 21 <sup>st</sup> day | $4.40 \pm 0.50^{a}$    | $5.00 \pm 0^{b}$      | $4.20 \pm 0.41^{a}$                              | $4.26 \pm 0.59^{a}$     | 0.001   |
|  | $28^{th} day$        | $5.00 \pm 0$           | $5.00 \pm 0$          | $5.00 \pm 0$                                     | $5.00 \pm 0$            | 1       |
| Color match (CM)                       | 3 <sup>rd</sup> day  | $4.33 \pm 4.87^{a,b}$  | $11.33 \pm 9.15^{a}$  | $3.33 \pm 6.17^{b}$                              | $6.66 \pm 8.17^{a,b}$   | 0.022   |
|  | $7^{th} day$         | $18.66 \pm 10.60^{a}$  | $40.66 \pm 10.99^{b}$ | $22.66 \pm 7.03^{a}$                             | $21.33 \pm 9.15^{a}$    | 0.001   |
|  | $14^{th} day$        | $59.33 \pm 7.98^{a}$   | $78.66 \pm 7.43^{b}$  | $60.66 \pm 10.99^{a}$                            | $58.00 \pm 6.76^{a}$    | 0.001   |
|  | 21 <sup>st</sup> day | $83.33 \pm 9.75^{a}$   | $95.33 \pm 5.16^{b}$  | $86.66 \pm 8.16^{a,b}$                           | $80.66 \pm 8.83^{a}$    | 0.001   |
|  | $28^{th} day$        | $96.00 \pm 5.07^{a,b}$ | $100 \pm 0^{a}$       | $96.66 \pm 4.87^{a,b} \qquad 95.33 \pm 5.16^{b}$ | 0.029                   |         |
| H <sub>2</sub> O <sub>2</sub> bubbling | 3 <sup>rd</sup> day  | $96.00 \pm 6.32$       | $96.00 \pm 6.32$      | $98.00 \pm 4.14$                                 | $97.33 \pm 5.93$        | 0.711   |
|  | $7^{th} day$         | $80.66 \pm 8.83^{a}$   | $69.33 \pm 10.32^{b}$ | $81.33 \pm 7.43^{a}$                             | $81.33 \pm 8.33^{a}$    | 0.004   |
|  | 14 <sup>th</sup> day | $12.00 \pm 9.41^{a,b}$ | $8.00 \pm 9.41^{a}$   | $21.33 \pm 11.87^{b}$                            | $18.66 \pm 12.45^{a,b}$ | 0.005   |
|  | 21 <sup>st</sup> day | $3.33 \pm 4.87$        | 0                     | $5.33 \pm 5.16$                                  | $4.66 \pm 5.16$         | 0.011   |
|  | $28^{th} day$        | 0                      | 0                     | 0  | 0                       | 1       |
| Analgesic consumption                  | 1 <sup>st</sup> day  | $2.26 \pm 0.70^{a}$    | $1.06\pm0.88^{\rm b}$ | $1.20 \pm 0.77^{b}$                              | $1.66 \pm 0.72^{b}$     | 0.001   |
|  | 3 <sup>rd</sup> day  | $2.20 \pm 0.67^{a}$    | $1.10\pm0.70^{\rm b}$ | $0.66 \pm 0.61^{b}$                              | $1.33 \pm 0.97^{\rm b}$ | 0.001   |
|  | 7 <sup>th</sup> day  | $0.33 \pm 0.61$        | $0.13 \pm 0.35$       | 0  | $0.35 \pm 0.62$         | 0.156   |

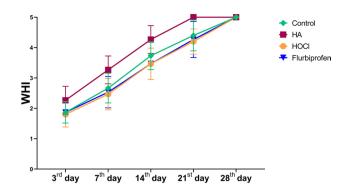


Fig. 2 Mean WHI values of groups

days. The chewing efficiency of the patients in the HOCl group was found to be significantly better in the flurbiprofen (p=0.001) and control groups (p=0.016) on the 1<sup>st</sup> day, and in the control group (p = 0.033) on the 3<sup>rd</sup> day. HOCl reduced the burning sensation compared to the flurbiprofen group (p=0.004) and the control group (p=0.039) on the 1<sup>st</sup> day. The burning sensation on the 3<sup>rd</sup> and 7<sup>th</sup> day in all study groups was found to be significantly lower than the control group, respectively (p < 0.05) (Table 3, Fig. 3).

HA group showed better CM with contiguous palatal tissue compared to the HOCl group (p = 0.037) on the 3<sup>rd</sup> day and those of all groups on the 7<sup>th</sup> and 14<sup>th</sup> days (p < 0.05). On day 21, the HA group had better results than the control (p = 0.003) and flurbiprofen (p = 0.001)groups, but on day 28, it achieved a better color match than flurbiprofen (p = 0.034) alone (Table 2, Fig. 4).

On the  $7^{\text{th}}$  day, the HA group showed less  $H_2O_2$  bubbling than the control (p=0.030), HOCl (p=0.015), and flurbiprofen groups (p = 0.015). On the 14<sup>th</sup> and 21<sup>st</sup> days, HA showed significant differences only with HOCl and flurbiprofen (p < 0.05). On the 28<sup>th</sup> day, epithelialization was completed in all groups (Table 2).

The photographs of palatal area were analyzed, and the CE was assessed visually. The HA group showed

| $3-8(5)^{a}$<br>$5-8(7)^{a}$<br>$3-6(5)^{a}$<br>0<br>0<br>$3-7(5)^{a}$<br>$2-7(4)^{a}$ | $2-8(5)^{a,b}$ $1-5(3)^{b,c,d}$ $0-3(1)^{b}$ $0$ $2-7(4)^{a,b}$ $0.5(2)^{a,b}$   | $\begin{array}{c} 0-6(3)^{b} \\ 0-3(2)^{c} \\ 0-1(0)^{b} \\ 0 \\ 0 \\ 0-7(2)^{b} \end{array}$ | $ \begin{array}{c} 1-7(5)^{a} \\ 1-6(3)^{d} \\ 0-4(1)^{b} \\ 0 \\ 0 \\ 3-7(5)^{a} \end{array} $   | 0.005<br>0.001<br>0.001<br>1<br>1  |
|--|--|---|---|--|
| 3-6(5) <sup>a</sup><br>0<br>0<br>3-7(5) <sup>a</sup>                                   | $0-3(1)^{b}$<br>0<br>0<br>$2-7(4)^{a,b}$   | 0–1(0) <sup>b</sup><br>0<br>0   | 04(1) <sup>b</sup><br>0<br>0  | <b>0.001</b><br>1<br>1   |
| 0<br>0<br>3-7(5) <sup>a</sup>  | $0-3(1)^{b}$<br>0<br>0<br>$2-7(4)^{a,b}$   | 0<br>0  | 04(1) <sup>b</sup><br>0<br>0  | 1<br>1   |
| 0<br>3–7(5) <sup>a</sup>   | 0 $2-7(4)^{a,b}$   | 0   | 0   | 1  |
| $3-7(5)^{a}$   | 2-7(4) <sup>a,b</sup>  |   |   |  |
|  |  | 0–7(2) <sup>b</sup>   | $3-7(5)^{a}$  | 0.001  |
| $2-7(4)^{a}$   |  |   |   | 0.001  |
|  | $0-5(3)^{a,b}$   | $0-4(2)^{b}$  | $1-6(3)^{a,b}$  | 0.021  |
| 0-2(1)   | 0-3(1)   | 0-2(0)  | 0–2(1)  | 0.134  |
| 0-2(0)   | 0  | 0   | 0   | 0.107  |
| 0  | 0  | 0   | 0   | 1  |
| $2-6(4)^{a}$   | $0-6(4)^{a,b}$   | $0-4(2)^{b}$  | $1-8(4)^{a}$  | 0.004  |
| $3-8(6)^{a}$   | $0-4(2)^{b}$   | $0-3(1)^{b}$  | $1-7(3)^{b}$  | 0.001  |
| $4-8(6)^{a}$   | $0-3(0)^{b}$   | $0^{b}$   | $0-5(0)^{b}$  | 0.001  |
| 0-1(0)   | 0  | 0   | 0-2(0)  | 0.106  |
| 0  | 0  | 0   | 0   | 1  |
|  | 0<br>2–6(4) <sup>a</sup><br>3–8(6) <sup>a</sup><br>4–8(6) <sup>a</sup><br>0–1(0) | $\begin{array}{llllllllllllllllllllllllllllllllllll$  | $\begin{array}{ccccc} 0 & 0 & 0 \\ 2-6(4)^{a} & 0-6(4)^{a,b} & 0-4(2)^{b} \\ 3-8(6)^{a} & 0-4(2)^{b} & 0-3(1)^{b} \\ 4-8(6)^{a} & 0-3(0)^{b} & 0^{b} \\ 0-1(0) & 0 & 0 \end{array}$ | $\begin{array}{cccccccc} 0 & 0 & 0 & 0 \\ 2-6(4)^{a} & 0-6(4)^{a,b} & 0-4(2)^{b} & 1-8(4)^{a} \\ 3-8(6)^{a} & 0-4(2)^{b} & 0-3(1)^{b} & 1-7(3)^{b} \\ 4-8(6)^{a} & 0-3(0)^{b} & 0^{b} & 0-5(0)^{b} \\ 0-1(0) & 0 & 0 & 0-2(0) \end{array}$ |

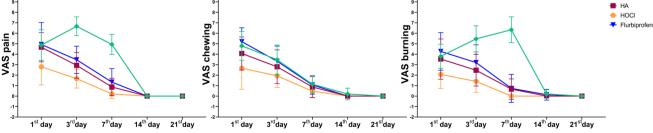


Fig. 3 Mean VAS pain, chewing, and burning values of groups

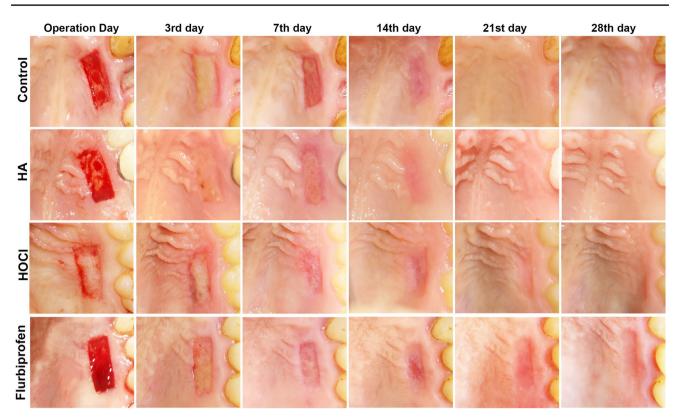


Fig. 4 In situ images of groups on the operation, 3<sup>rd</sup>, 7<sup>th</sup>, 14<sup>th</sup>, 21<sup>st</sup>, and 28<sup>th</sup> days

| Parameters                   | Days                 |     | Control $n=15$                         | $ \begin{array}{c} \text{HA} \\ n = 15 \end{array} $ | HOCl<br>n=15          | Flurbiprofen $n = 15$ | <i>p</i> value |
|------------------------------|----------------------|-----|--|--|-----------------------|-----------------------|----------------|
| Complete epithelization (CE) | 3 <sup>rd</sup> day  | No  | 15(100%)                               | 15(100%)   | 15(100%)              | 15(100%)              |                |
|                              | 7 <sup>th</sup> day  | No  | 15(100%)                               | 15(100%)   | 15(100%)              | 15(100%)              |                |
|                              | 14 <sup>th</sup> day | No  | 11(73.3%)                              | 7(46.7%)   | 12(80%)               | 11(73.3%)             | 0.054          |
|                              |                      | Yes | 4(26.7%)                               | 8(53.3%)   | 3(20%)                | 4(26.7%)              |                |
|                              | 21 <sup>th</sup> day | No  | 5(33.3%) <sup>a,b</sup>                | 0(0) <sup><b>a</b></sup>                             | 8(53.3%) <sup>b</sup> | 7(46.7%) <sup>b</sup> | 0.010          |
|                              |                      | Yes | 10(66.7%) <sup><b>a</b>,<b>b</b></sup> | 15(100%) <sup>a</sup>                                | 7(46.7%) <sup>b</sup> | 8(53.3%) <sup>b</sup> |                |
|                              | $28^{th} day$        | Yes | 15(100%)                               | 15(100%)   | 15(100%)              | 15(100%)              |                |
| Delayed bleeding (DB)        | 1 <sup>st</sup> day  | No  | 8(53.3%)                               | 12(80%)  | 7(46.7%)              | 11(73.3%)             | 0.181          |
|                              |                      | Yes | 7(46.7%)                               | 3(20%)   | 8(53.3%)              | 4(26.7%)              |                |
|                              | 3 <sup>rd</sup> day  | No  | 10(66.7%)                              | 14(93.3%)  | 10(66.7%)             | 12(80%)               | 0.200          |
|                              |                      | Yes | 5(33.3%)                               | 1(6.7%)  | 5(33.3%)              | 3(20%)                |                |
|                              | 7 <sup>th</sup> day  | No  | 12(80%)                                | 15(100%)   | 13(86.7%)             | 13(86.7%)             | 0.199          |
|                              |                      | Yes | 3(20%)                                 | 0(0)   | 2(13.3%)              | 2(13.3%)              |                |

Table 4 Complete epithelization (CE) and delayed bleeding (DB) values of the groups. Data were presented as count (% of total). p < 0.05; the significant difference between groups with the different superscripts

significantly better epithelialization than the HOCL and flurbiprofen groups on the  $21^{st}$  day (Fig. 4, Table 4). There was no difference between the groups in terms of delayed bleeding on the day of the  $1^{st}$ ,  $3^{rd}$ , and  $7^{th}$  days postoperatively (Table 4, Fig. 4).

# Discussion

The most unfavorable condition after FGG surgery for patients is palatal donor site morbidity. Numerous clinical studies have focused on enhancing palatal wound healing and reducing patient discomfort [8, 26, 32, 33]. However, there is no study in the literature evaluating the effects of HOCl on palatal wound healing after periodontal plastic surgery. The present randomized clinical study was designed to investigate the therapeutic effects of different topical agents on the secondary wound healing of the donor palatal region and the patient's discomfort after the FGG procedure. The primary study outcome (WHI) indicated that the HA group revealed the best healing scores up to 21 days postoperatively. Concomitantly, higher rates of CE on the 14<sup>th</sup> day were observed in the HA group compared to the other study groups. However, HOCL resulted in lower VAS levels for the pain, chewing, and burning sensation parameters compared to the controls.

Many steps are necessary for wound healing to take place, among them; cell differentiation, proliferation, migration, and collagen deposition [34, 35]. HA is a member of GAGs, which is the main component of the extracellular matrix (ECM) and has beneficial effects on wound healing by reducing inflammation, increasing vascularization, and collagen synthesis [36-38]. HA is involved in all steps of the woundhealing process [39]. Yıldırım et al. [26] compared two different high molecular weight HA gels (0.2% and 0.8%) in terms of donor site healing. According to the results, healing was found to be better in the 0.2% gel group on the  $14^{\text{th}}$  day, while both HA groups showed better epithelialization on the 21<sup>st</sup> day than the control. Chen et al. [40] reported that high molecular weight HA when combined with povidone-iodine, significantly improved wound healing, and promoted both cell proliferation and neovascularization at the wound site compared to low molecular weight HA. Increased fibroblast proliferation, rapid wound closure, and increased inflammatory cell infiltration have been reported after the topical application of HA [41]. In addition, in an animal study, it was found that the use of HA in the wound healing model in groups treated with HA increased the levels of fibroblast, collagen I, and collagen III consequently accelerating wound healing [42]. There are studies indicating that HA stimulates keratinocyte migration and proliferation and has a positive effect on re-epithelialization [43]. Since HA plays a curative role in every step of wound healing, we also obtained the best wound healing scores in the groups treated with HA in our study, and complete epithelialization was achieved in the HA group in 21 days compared to the other groups.

In our study, VAS scores on the  $3^{rd}$  and  $7^{th}$  days were found to be lower in the HA group compared to the control group, and analgesic consumption of patients was lower in the HA group on the  $1^{st}$  and  $7^{th}$  days compared to the control group. After FGG surgery, locally applied HA gel (a mixture of cross-linked (1,6%) and natural (0,2%) HA) was compared with the control group and followed for six months. According to the results of the study, there was no difference between the two groups in terms of CM, and pain levels were higher in the control group compared to the HA group in the first seven days [44]. Hassan et al. [45] compared 0.2% HA gel, MEBO, and the control group, and it was found that the study groups showed significantly lower VAS scores on the 2<sup>nd</sup> and 3<sup>rd</sup> days compared to the control group. They stated that from the 4th day, the patients did not have any pain although there was no difference between the groups in terms of wound size at any time. These differences may be due to differences in HA molecular weight and concentration. In our study, we used high molecular weight HA at 0.6% concentration. High molecular weight HA exhibits anti-inflammatory and immunosuppressive properties. It has been reported that high molecular weight HA inhibited the IL-1β expression in a rabbit osteoarthritis study [46]. In an osteoarthritis model, high molecular weight HA downregulated IL-8 and the inducible nitrous oxide synthase gene expression and downregulated TNF- $\alpha$ gene expression in IL-1-stimulated fibroblast-like synoviocytes [47]. Campo et al. [48] found a reduction of mRNA expression and protein production for TNF-α, IL-1β, IL-17, matrix metalloproteinases-13, and the inducible nitrous oxide synthase gene in high molecular weight HA-treated arthritic mice. Furthermore high molecular weight HA was found to decrease the IL-1 $\beta$ , IL-6, TNF- $\alpha$ , and nitric oxide in microglial cells exposed to lipopolysaccharide [49]. Considering the necessity of increasing certain cytokines for the formation of acute pain, HA could decrease the perception of pain in patients by these mechanisms.

As the risks of infectious diseases increase around the world, the need for an effective broad-spectrum antimicrobial agent is increasingly becoming a therapeutic imperative. Because it inactivates the SARS-CoV-2 virus, HOCl has become popular in dentistry as a disinfectant and antiseptic during the COVID-19 pandemic [50, 51]. Naturally, respiratory bursts during the activation of neutrophils produce peroxide  $(H_2O_2)$  and the activated granule enzyme myeloperoxidase converts H<sub>2</sub>O<sub>2</sub> to HOCl in the presence of  $Cl^{-}$  and  $H^{+}$  [52]. The resulting HOCl contributes to the bactericidal activity of neutrophils and is thought to cause tissue damage in areas of inflammation. In many studies, it has been stated that the use of HOCl has positive effects on wound healing [53]. In an in-vitro study, the effectiveness of NaClO/HClO solutions on wound healing was investigated and an increased antimicrobial effect was associated with decreased viability of keratinocytes and fibroblasts. The authors noted that the microbicidal effects almost always have a certain negative impact on cell proliferation and viability [54]. HOCl is a weak acid and an oxidant and also has high interaction potential with other molecules in a redox reaction, resulting in the formation of reactive oxygen species (ROS: hydrogen peroxide, superoxide, hydroxyl radicals, oxygen) [54, 55]. Excessive ROS may alter and/ or degrade ECM proteins, resulting in impaired dermal

fibroblast and keratinocyte function [56]. Considering the results of our study, lower WHI scores were obtained in the HOCl-administered group compared to the other groups, including the controls, in all time periods. Studies have also emphasized that HOCl is a selective oxidant that can easily react with cellular proteins like fibronectin, thrombospondin, and laminin, and may also cause extracellular matrix fragmentation and tissue denaturation [57, 58]. The resulting protein damage was found to be related to the amount of HOCl administered rather than the concentration [54, 58]. This result can be explained as follows: HOCl application causes an increased ROS in the palatal region and may result in a delay in wound healing. When HOCl and saline were compared in a histological study, although the number of methicillin-resistant Staphylococcus aureus (MRSA) was found to be decreased in the HOCl group compared to the control, there was no difference between the two groups in terms of epithelial thickness and granulation tissue formation [59]. In a randomized controlled trial (RCT) performed on the acute wound model, the use of HOCl was beneficial for epithelialization in the first days, but no difference was found in terms of re-epithelialization versus saline use on day 10. In addition, there was no significant difference between the two groups in VAS pain scores [60]. Mekkawy et al. [61] investigated the efficacy of HOCl on septic traumatic wounds in an RCT and found lower VAS scores up to 14 days than the control group, consistent with our study. In our study, pain, chewing, and burning scores evaluated by VAS in the HOCl group were found to be significantly lower than the control group up to 7 days. The side effect of a bacterial infection can be a tender, painful wound [62]. Proinflammatory cytokines such as ILs, TNF- $\alpha$ , granulocyte-macrophage colony-stimulating factor (GM-CSF), and monocyte chemoattractant protein-1 may be potential biomarkers for acute surgical wound pain [63]. HOCl has anti-inflammatory and immunomodulatory properties and reduces histamine, leukotriene B4, IL-6, and IL-2 activities [64]. Some authors have suggested that super-oxidized solutions act as a mast cell membrane stabilizing inhibitor, and diminish the mast-cell degranulation induced by IgE-antigen receptor cross-linking [65]. The fact that the VAS scores in our study were lower in the HOCl-administered groups than in the other groups may be the result of the suppression of inflammation due to the effects of HOCl, an antibacterial agent, on cytokine expression.

Flurbiprofen decreases prostaglandin synthesis by inhibiting the COX-2 enzyme induced immediately in response to injury [66] but prostaglandins organize the cellular proliferation, vascular permeability, and angiogenesis, involved in wound healing [67]. In an animal model, COX-2 inhibitors caused delayed re-epithelialization in excisional wounds at cutaneous tissue [68]. Additionally, it was reported that COX-2 selective inhibitors resulted in significant inhibition of angiogenesis [69]. In wounded gastric epithelial cells, Pai et al. indicated that NSAIDs decrease both basal and epidermal growth factor induced re-epithelialization [70]. In our study, mean WHI values in the flurbiprofen group were similar for the control and HOCL groups at all-time points, but significantly lower than the HA group. Although the prevalence of CE was lower on the 21st day compared to the control group, it was not statistically significant. Isler et al. investigated the effects of topical flurbiprofen on the palatal donor site after both subgingival connective tissue graft (SCGT) and FGG procedures. According to their results, flurbiprofen resulted in delayed epithelization on the 21st day compared to the placebo group, and epithelization was completed within 42 days for all patients [33]. Contrary to these results, the prevalence of CE was 100% in 28 days in our study. It was shown that residual tissue thickness at the donor site after FGG harvesting affects the speed of palatal wound filling directly [71]. The lower graft thickness obtained in the present study may have resulted in faster re-epithelization in the palatal donor area. Many studies show that topically administered NSAIDs are effective in the management of postoperative pain after tonsillectomy [10, 72]. Koray et al. found similar VAS pain scores of NSAI spray (benzydamine hydrochloride) application compared to hyaluronic acid spray (0.2%) after third molar surgery [73]. In the study of Isler et.al, flurbiprofen spray decreased the VAS pain levels compared to the placebo group throughout the study period [33]. Consistent with the results of these studies, in the present study, VAS scores in the flurbiprofen group were similar HA group but were lower than controls on the 3<sup>rd</sup> and 7<sup>th</sup> days postoperatively.

The present study had some limitations. VAS provides a subjective evaluation of the pain, chewing, and burning sensation rather than an objective parameter. Although the VAS scores showed a statistically significant difference between the groups at the time of evaluation, there was no difference between the groups in terms of analgesic consumption by the patients. Despite the statistically significant differences detected between groups, they may only be of very low clinical relevance. Also, using a locally applied therapeutic agent by patients may have caused them to report lower VAS scores in study groups compared to the controls. Another limitation of our study is the inability to examine cytokine expression by histological examination due to ethical barriers.

# Conclusion

Clinicians should consider possible beneficial effects on the secondary wound healing process and patients' discomfort; high molecular weight HA may be the first choice for the management of palatal donor site morbidity after FGG procedures. Although the use of HOCl and flurbiprofen reduced the pain compared to the control group, delayed epithelialization was observed due to possible effects on the inflammatory phase of wound healing.

Author contribution Both authors have contributed equally to the work.

**Data Availability** The data that support the findings of this study are available from the corresponding author, upon reasonable request.

#### Declarations

Competing interests The authors declare no competing interests.

**Ethics approval** Permission was obtained from the Ethics Committee of Pamukkale University (05.03.2019/05) for the study protocol.

Conflict of interest The authors declare no conflict of interest.

## References

- Camargo PM, Melnick PR, Kenney EB (2001) The use of free gingival grafts for aesthetic purposes. Periodontol 2000 27:72–96. https://doi.org/10.1034/j.1600-0757.2001.027001072.x
- 2. Farnoush A (1978) Techniques for the protection and coverage of the donor sites in free soft tissue grafts. J Periodontol 49:403–405. https://doi.org/10.1902/jop.1978.49.8.403
- Burkhardt R, Hammerle CH, Lang NP, Research Group on Oral Soft Tissue B, Wound H (2015) Self-reported pain perception of patients after mucosal graft harvesting in the palatal area. J Clin Periodontol 42:281–7. https://doi.org/10.1111/jcpe.12357
- Griffin TJ, Cheung WS, Zavras AI, Damoulis PD (2006) Postoperative complications following gingival augmentation procedures. J Periodontol 77:2070–2079. https://doi.org/10.1902/jop. 2006.050296
- Keceli HG, Aylikci BU, Koseoglu S, Dolgun A (2015) Evaluation of palatal donor site haemostasis and wound healing after free gingival graft surgery. J Clin Periodontol 42:582–589. https://doi. org/10.1111/jcpe.12404
- Roccuzzo A, Imber JC, Bosshardt D, Salvi GE, Sculean A (2021) Development of bone exostosis following the use of a free gingival graft: a 30-year case report and literature review. Int J Periodont Restor Dent 41:539–545. https://doi.org/10.11607/prd.5035
- Yussif N, Wagih R, Selim K (2021) Propylene mesh versus acrylic resin stent for palatal wound protection following free gingival graft harvesting: a short-term pilot randomized clinical trial. BMC Oral Health 21:208. https://doi.org/10.1186/s12903-021-01541-z
- Ustaoglu G, Ercan E, Tunali M (2016) The role of titanium-prepared platelet-rich fibrin in palatal mucosal wound healing and histoconduction. Acta Odontol Scand 74:558–564. https://doi.org/ 10.1080/00016357.2016.1219045
- Kozlovsky A, Artzi Z, Hirshberg A, Israeli-Tobias C, Reich L (2007) Effect of local antimicrobial agents on excisional palatal wound healing: a clinical and histomorphometric study in rats. J Clin Periodontol 34:164–171. https://doi.org/10.1111/j.1600-051X.2006.01033.x
- Turk B, Akpinar M, Erol ZN, Kaya KS, Unsal O, Coskun BU (2018) The effect of flurbiprofen oral spray and ibuprofen vs ibuprofen alone on postoperative tonsillectomy pain: an open,

randomised, controlled trial. Clin Otolaryngol 43:835–840. https://doi.org/10.1111/coa.13058

- Muderris T, Tezcan G, Sancak M, Gul F, Ugur G (2019) Oral flurbiprofen spray for postoperative sore throat and hoarseness: a prospective, randomized, double-blind, placebo-controlled study. Minerva Anestesiol 85:21–27. https://doi.org/10.23736/S0375-9393.18.12703-9
- Ooki A, Del Carmen Rodriguez Pena M, Marchionni L, Dinalankara W, Begum A, Hahn NM, VandenBussche CJ, Rasheed ZA, Mao S, Netto GJ, Sidransky D, Hoque MO (2018) YAP1 and COX2 coordinately regulate urothelial cancer stem-like cells. Cancer Res 78:168–181. https://doi.org/10.1158/0008-5472. CAN-17-0836
- Aich A, Wang C, Chowdhury A, Ronsor C, Pacheu-Grau D, Richter-Dennerlein R, Dennerlein S, Rehling P (2018) COX16 promotes COX2 metallation and assembly during respiratory complex IV biogenesis. Elife 7. https://doi.org/10.7554/eLife. 32572
- Jiang WW, Wang QH, Peng P, Liao YJ, Duan HX, Xu M, Li Y, Zhang PB (2015) Effects of flurbiprofen axetil on postoperative serum IL-2 and IL-6 levels in patients with colorectal cancer. Genet Mol Res 14:16469–16475. https://doi.org/10.4238/2015. December.9.18
- Wang X, Ye X, Zhang Y, Ji F (2020) Flurbiprofen suppresses the inflammation, proliferation, invasion and migration of colorectal cancer cells via COX2. Oncol Lett 20:132. https://doi.org/10. 3892/ol.2020.11993
- 16. Robson MC, Payne WG, Ko F, Mentis M, Donati G, Shafii SM, Culverhouse S, Wang L, Khosrovi B, Najafi R, Cooper DM, Bassiri M (2007) Hypochlorous acid as a potential wound care agent: part II. stabilized hypochlorous acid: its role in decreasing tissue bacterial bioburden and overcoming the inhibition of infection on wound healing. J Burns Wounds 6:e6
- 17. Gold MH, Andriessen A, Bhatia AC, Bitter P Jr, Chilukuri S, Cohen JL, Robb CW (2020) Topical stabilized hypochlorous acid: the future gold standard for wound care and scar management in dermatologic and plastic surgery procedures. J Cosmet Dermatol 19:270–277. https://doi.org/10.1111/jocd.13280
- Wang L, Bassiri M, Najafi R, Najafi K, Yang J, Khosrovi B, Hwong W, Barati E, Belisle B, Celeri C, Robson MC (2007) Hypochlorous acid as a potential wound care agent: part I. Stabilized hypochlorous acid: a component of the inorganic armamentarium of innate immunity. J Burns Wounds 6:e5
- Graca MFP, Miguel SP, Cabral CSD, Correia IJ (2020) Hyaluronic acid-based wound dressings: a review. Carbohydr Polym 241:116364. https://doi.org/10.1016/j.carbpol.2020.116364
- 20. Gencer ZK, Ozkiris M, Okur A, Korkmaz M, Saydam L (2014) A comparative study on the impact of intra-articular injections of hyaluronic acid, tenoxicam and betametazon on the relief of temporomandibular joint disorder complaints. J Craniomaxillofac Surg 42:1117–1121. https://doi.org/10.1016/j.jcms.2014.01.041
- Prosdocimi M, Bevilacqua C (2012) Exogenous hyaluronic acid and wound healing: an updated vision. Panminerva Med 54:129–135
- 22. Voigt J, Driver VR (2012) Hyaluronic acid and wound healing. Wound Repair Regen 20:317–331. https://doi.org/10.1111/j.1524-475X.2012.00777.x
- Voigt J, Driver VR (2012) Hyaluronic acid derivatives and their healing effect on burns, epithelial surgical wounds, and chronic wounds: a systematic review and meta-analysis of randomized controlled trials. Wound Repair Regen 20:317–331. https://doi. org/10.1111/j.1524-475X.2012.00777.x
- Neuman MG, Nanau RM, Oruna-Sanchez L, Coto G (2015) Hyaluronic acid and wound healing. J Pharm Pharm Sci 18:53–60. https://doi.org/10.18433/j3k89d

- Humbert P, Mikosinki J, Benchikhi H, Allaert FA (2013) Efficacy and safety of a gauze pad containing hyaluronic acid in treatment of leg ulcers of venous or mixed origin: a double-blind, randomised, controlled trial. Int Wound J 10:159–166. https://doi. org/10.1111/j.1742-481X.2012.00957.x
- Yildirim S, Ozener HO, Dogan B, Kuru B (2018) Effect of topically applied hyaluronic acid on pain and palatal epithelial wound healing: an examiner-masked, randomized, controlled clinical trial. J Periodontol 89:36–45. https://doi.org/10.1902/jop.2017. 170105
- Bergstrand S, Ingstad HK, Moystad A, Bjornland T (2019) Longterm effectiveness of arthrocentesis with and without hyaluronic acid injection for treatment of temporomandibular joint osteoarthritis. J Oral Sci 61:82–88. https://doi.org/10.2334/josnusd. 17-0423
- Iturriaga V, Vasquez B, Bornhardt T, Del Sol M (2021) Effects of low and high molecular weight hyaluronic acid on the osteoarthritic temporomandibular joint in rabbit. Clin Oral Investig 25:4507–4518. https://doi.org/10.1007/s00784-020-03763-x
- Günaydın B (2021) ASA Fiziksel Durum Sınıflandırma Sistemi: ASA Physical Status Classification System. Turk J Anaesthesiol Reanim 49:192–193
- Guerrero A, Griffiths GS, Nibali L, Suvan J, Moles DR, Laurell L, Tonetti MS (2005) Adjunctive benefits of systemic amoxicillin and metronidazole in non-surgical treatment of generalized aggressive periodontitis: a randomized placebo-controlled clinical trial. J Clin Periodontol 32:1096–1107. https://doi.org/10.1111/j. 1600-051X.2005.00814.x
- Landry RG (1985) Effectiveness of benzydamine HC1 in the treatment of periodontal post-surgical patients. Faculty of Dentistry, University of Toronto
- Lektemur Alpan A, Torumtay Cin G (2020) PRF improves wound healing and postoperative discomfort after harvesting subepithelial connective tissue graft from palate: a randomized controlled trial. Clin Oral Investig 24:425–436. https://doi.org/10.1007/ s00784-019-02934-9
- Isler SC, Eraydin N, Akkale H, Ozdemir B (2018) Oral flurbiprofen spray for mucosal graft harvesting at the palatal area: a randomized placebo-controlled study. J Periodontol 89:1174–1183. https://doi.org/10.1002/jper.17-0381
- Broughton G 2nd, Janis JE, Attinger CE (2006) The basic science of wound healing. Plast Reconstr Surg 117:12S-34S. https://doi. org/10.1097/01.prs.0000225430.42531.c2
- Velnar T, Bailey T, Smrkolj V (2009) The wound healing process: an overview of the cellular and molecular mechanisms. J Int Med Res 37:1528–1542. https://doi.org/10.1177/147323000903700531
- Aya KL, Stern R (2014) Hyaluronan in wound healing: rediscovering a major player. Wound Repair Regen 22:579–593. https://doi. org/10.1111/wrr.12214
- 37. Frenkel JS (2014) The role of hyaluronan in wound healing. Int Wound J 11:159–163. https://doi.org/10.1111/j.1742-481X.2012. 01057.x
- Kasuya A, Tokura Y (2014) Attempts to accelerate wound healing. J Dermatol Sci 76:169–172. https://doi.org/10.1016/j.jderm sci.2014.11.001
- Litwiniuk M, Krejner A, Speyrer MS, Gauto AR, Grzela T (2016) Hyaluronic acid in inflammation and tissue regeneration. Wounds 28:78–88
- Chen RF, Wang CT, Chen YH, Chien CM, Lin SD, Lai CS, Wang CJ, Kuo YR (2019) Hyaluronic acid-povidone-iodine compound facilitates diabetic wound healing in a streptozotocin-induced diabetes rodent model. Plast Reconstr Surg 143:1371–1382. https:// doi.org/10.1097/prs.00000000005504
- 41. Tolg C, Telmer P, Turley E (2014) Specific sizes of hyaluronan oligosaccharides stimulate fibroblast migration and excisional

wound repair. PLoS ONE 9:e88479. https://doi.org/10.1371/journ al.pone.0088479

- 42. Taskan MM, Balci Yuce H, Karatas O, Gevrek F, Isiker Kara G, Celt M, Sirma Taskan E (2021) Hyaluronic acid with antioxidants improve wound healing in rats. Biotechnol Histochem 96:536–545. https://doi.org/10.1080/10520295.2020.1832255
- Nyman E, Henricson J, Ghafouri B, Anderson CD, Kratz G (2019) Hyaluronic acid accelerates re-epithelialization and alters protein expression in a human wound model. Plast Reconstr Surg Glob Open 7:e2221–e2221. https://doi.org/10.1097/GOX.000000000 002221
- 44. Khalil S, Habashneh RA, Alomari S, Alzoubi M (2022) Local application of hyaluronic acid in conjunction with free gingival graft: a randomized clinical trial. Clin Oral Invest 26:2165–2174. https://doi.org/10.1007/s00784-021-04197-9
- 45. Hassan A, Ahmed E, Ghalwash D, Elarab AE (2021) Clinical comparison of MEBO and hyaluronic acid gel in the management of pain after free gingival graft harvesting: a randomized clinical trial. Int J Dent 2021:2548665–2548665. https://doi.org/10.1155/ 2021/2548665
- 46. Takahashi K, Goomer RS, Harwood F, Kubo T, Hirasawa Y, Amiel D (1999) The effects of hyaluronan on matrix metalloproteinase-3 (MMP-3), interleukin-1beta(IL-1beta), and tissue inhibitor of metalloproteinase-1 (TIMP-1) gene expression during the development of osteoarthritis. Osteoarthritis Cartil 7:182–190. https://doi.org/10.1053/joca.1998.0207
- 47. Wang CT, Lin YT, Chiang BL, Lin YH, Hou SM (2006) High molecular weight hyaluronic acid down-regulates the gene expression of osteoarthritis-associated cytokines and enzymes in fibroblast-like synoviocytes from patients with early osteoarthritis. Osteoarthritis Cartil 14:1237–1247. https://doi.org/10.1016/j. joca.2006.05.009
- Campo GM, Avenoso A, Nastasi G, Micali A, Prestipino V, Vaccaro M, D'Ascola A, Calatroni , Campo S (2011) Hyaluronan reduces inflammation in experimental arthritis by modulating TLR-2 and TLR-4 cartilage expression. Biochim Biophysica Acta (BBA) Mol Basis Dis 1812:1170–1181. https://doi.org/10.1016/j. bbadis.2011.06.006
- Austin JW, Gilchrist C, Fehlings MG (2012) High molecular weight hyaluronan reduces lipopolysaccharide mediated microglial activation. J Neurochem 122:344–355. https://doi.org/10. 1111/j.1471-4159.2012.07789.x
- Block MS, Rowan BG (2020) Hypochlorous acid: a review. J Oral Maxillofac Surg 78:1461–1466. https://doi.org/10.1016/j.joms. 2020.06.029
- Hatanaka N, Yasugi M, Sato T, Mukamoto M, Yamasaki S (2022) Hypochlorous acid solution is a potent antiviral agent against SARS-CoV-2. J Appl Microbiol 132:1496–1502. https://doi.org/ 10.1111/jam.15284
- Vissers MC, Winterbourn CC (1995) Oxidation of intracellular glutathione after exposure of human red blood cells to hypochlorous acid. Biochem J 307(Pt 1):57–62. https://doi.org/10.1042/ bj3070057
- Joachim D (2020) Wound cleansing: benefits of hypochlorous acid. J Wound Care 29:S4–S8. https://doi.org/10.12968/jowc. 2020.29.Sup10a.S4
- Severing A-L, Rembe J-D, Koester V, Stuermer EK (2018) Safety and efficacy profiles of different commercial sodium hypochlorite/ hypochlorous acid solutions (NaClO/HClO): antimicrobial efficacy, cytotoxic impact and physicochemical parameters in vitro. J Antimicrob Chemother 74:365–372. https://doi.org/10.1093/jac/ dky432
- 55. Armstrong DG, Bohn G, Glat P, Kavros SJ, Kirsner R, Snyder R, Tettelbach W (2015) Expert recommendations for the use of hypochlorous solution: science and clinical application. Ostomy Wound Manag 61:S2-s19

- 56. Moseley R, Stewart JE, Stephens P, Waddington RJ, Thomas DW (2004) Extracellular matrix metabolites as potential biomarkers of disease activity in wound fluid: lessons learned from other inflammatory diseases? Br J Dermatol 150:401–413. https://doi.org/10. 1111/j.1365-2133.2004.05845.x
- Woods AA, Davies MJ (2003) Fragmentation of extracellular matrix by hypochlorous acid. Biochem J 376:219–227. https:// doi.org/10.1042/BJ20030715
- Vissers MCM, Thomas C (1997) Hypochlorous acid disrupts the adhesive properties of subendothelial matrix. Free Radic Biol Med 23:401–411. https://doi.org/10.1016/S0891-5849(96) 00619-3
- 59. Davis SC, Gil J, Li J, Simms C, Valdes J, Solis M, Higa A (2021) Effect of mechanical debridement and irrigation with hypochlorous acid wound management solution on methicillin-resistant Staphylococcus aureus contamination and healing deep dermal wounds in a porcine model. Wound Manag Prev 67:24–31
- Burian EA, Sabah L, Kirketerp-Møller K, Gundersen G, Ågren MS (2022) Effect of stabilized hypochlorous acid on re-epithelialization and bacterial bioburden in acute wounds: a randomized controlled trial in healthy volunteers. Acta Dermato-Venereol 102:adv00727. https://doi.org/10.2340/actady.v102.1624
- Mekkawy MM, Kamal A (2014) A randomized clinical trial: the efficacy of hypochlorous acid on septic traumatic wound. J Educ Pract 5:89–100
- Zhao G, Usui ML, Lippman SI, James GA, Stewart PS, Fleckman P, Olerud JE (2013) Biofilms and inflammation in chronic wounds. Adv Wound Care (New Rochelle) 2:389–399. https://doi. org/10.1089/wound.2012.0381
- Goto T, Saligan LN (2020) Wound pain and wound healing biomarkers from wound exudate: a scoping review. J Wound Ostomy Continence Nurs 47:559–568. https://doi.org/10.1097/won.00000 00000000703
- Del Rosso JQ, Bhatia N (2018) Status report on topical hypochlorous acid: clinical relevance of specific formulations, potential modes of action, and study outcomes. J Clin Aesthet Dermatol 11:36–39
- 65. Medina-Tamayo J, Sánchez-Miranda E, Balleza-Tapia H, Ambriz X, Cid ME, González-Espinosa D, Gutiérrez AA, González-Espinosa C (2007) Super-oxidized solution inhibits IgE-antigeninduced degranulation and cytokine release in mast cells. Int Immunopharmacol 7:1013–1024. https://doi.org/10.1016/j.intimp. 2007.03.005

- Davies NM (1995) Clinical pharmacokinetics of flurbiprofen and its enantiomers. Clin Pharmacokinet 28:100–114. https://doi.org/ 10.2165/00003088-199528020-00002
- Ricciotti E, FitzGerald GA (2011) Prostaglandins and inflammation. Arterioscler Thromb Vasc Biol 31:986–1000. https://doi.org/ 10.1161/ATVBAHA.110.207449
- Fairweather M, Heit YI, Buie J, Rosenberg LM, Briggs A, Orgill DP, Bertagnolli MM (2015) Celecoxib inhibits early cutaneous wound healing. J Surg Res 194:717–724. https://doi.org/10.1016/j. jss.2014.12.026
- 69. Masferrer JL, Koki A, Seibert K (1999) COX-2 inhibitors. A new class of antiangiogenic agents. Ann N Y Acad Sci 889:84–86. https://doi.org/10.1111/j.1749-6632.1999.tb08726.x
- Pai R, Szabo IL, Giap AQ, Kawanaka H, Tarnawski AS (2001) Nonsteroidal anti-inflammatory drugs inhibit re-epithelialization of wounded gastric monolayers by interfering with actin, Src, FAK, and tensin signaling. Life Sci 69:3055–3071. https://doi. org/10.1016/s0024-3205(01)01412-6
- Keskiner I, Aydogdu A, Balli U, Kaleli AE (2016) Quantitative changes in palatal donor site thickness after free gingival graft harvesting: a pilot study. J Clin Periodontol 43:976–984. https:// doi.org/10.1111/jcpe.12592
- Muderris T, Gul F, Yalciner G, Babademez MA, Bercin S, Kiris M (2016) Oral flurbiprofen spray for posttonsillectomy pain. Otolaryngol Head Neck Surg 155:166–172. https://doi.org/10.1177/ 0194599816637865
- Koray M, Ofluoglu D, Onal EA, Ozgul M, Ersev H, Yaltirik M, Tanyeri H (2014) Efficacy of hyaluronic acid spray on swelling, pain, and trismus after surgical extraction of impacted mandibular third molars. Int J Oral Maxillofac Surg 43:1399–1403. https:// doi.org/10.1016/j.ijom.2014.05.003

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