

Effects of Intravenous and Intracerebroventricular Theophylline on Hypoxic Ventilatory Depression in Anesthetized Cats

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Key Words

Moderate hypoxia · Hypoxic depression of ventilation · Theophylline

Abstract

Objective: The present study was undertaken to investigate the ventilatory response due to sustained isocapnic moderate hypoxia and the possible role of adenosine in hypoxic depression in anesthetized cats. **Materials and Methods:** Cats anesthetized with pentothal sodium (30 mg kg⁻¹ i.p.) were divided into two groups: treated (n = 11) and control (n = 15). Respiratory frequency (f), tidal volume (V_T), minute volume (VE) and systemic arterial blood pressure were recorded during air and 20 min of breathing hypoxic gas mixture (14% O₂-86% N₂). Isocapnia was maintained by adding fractions of 1% CO₂ to the inspired hypoxic gas mixture. The PaO₂ and PaCO₂ were determined. **Results:** On hypoxic gas mixture breathing, V_T and VE values of the control animals increased significantly, at 5 min to 50 ± 6 and 53 ± 6%, respectively, above the prehypoxic air phase value (p < 0.001). After that, the magnitude of increase in V_T and VE declined gradually. At 20 min of hypoxia, V_T and VE were less than those in prehypoxic air phase (17 ± 7, 16 ± 7%, respectively). In cats injected with an adenosine antagonist

(theophylline 13.6 mg kg⁻¹ i.v.), f, V_T and VE increased significantly at 5 min of hypoxia (p < 0.001). At 20 min of hypoxia, f, V_T and VE were 8 ± 2, 30 ± 8, and 39 ± 8%, respectively, higher than corresponding values of the prehypoxic stage. In cats injected with theophylline (0.5 mg kg⁻¹) by cisternal puncture V_T and VE increased significantly at 5 min of hypoxia. At 20 min of hypoxia, V_T and VE were 27 ± 7 and 31 ± 8% higher than those in the prehypoxic air phase. **Conclusion:** The results of this study show that accumulation of adenosine in the brain during hypoxia seems to reduce the response of the central mechanisms to chemoreceptor impulses.

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Introduction

The ventilatory response to isocapnic sustained hypoxia is biphasic [1, 2]. A rapid increase in ventilation is followed by a decrease in respiratory minute volume [1–3]. The initial rise in ventilation is due to stimulation of the peripheral chemoreceptors. The secondary decrease is thought to be of central origin [1, 2] and has been termed ‘hypoxic depression’ or ‘roll-off’ [2, 4, 5]. Hypoxic depression of ventilation is absent when the brain stem is kept hyperoxic by artificial perfusion [1]. It is well known that

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peripheral chemoreceptors mediate the hyperventilatory response to hypoxia. In the absence of peripheral chemoreceptor inputs, hypoxia does not produce a hyperventilatory response. It has been clearly shown that hypoxia depresses ventilation in peripherally chemodenervated animals with or without vagotomy due to its central depressant effect [6–8]. Direct evidence of a central depressant effect of hypoxia was also obtained from artificial perfusion studies in which hypoxia was exclusively applied to the brain stem [9, 10]. In these studies the pons, medulla oblongata and cerebellum of anesthetized cats were perfused artificially with blood in which blood gas tensions varied independently from those in the systemic circulation hence leading to a separation of the central and peripheral effect of O₂ tension on ventilation. The studies showed that the central depression of ventilation was caused by decreases in CO₂ tension at the central chemosensors during hypoxia induced by increases in cerebral blood flow and Haldane effect [10, 11]. Artificial perfusion studies have also shown that the functioning of respiratory neurons in the brain stem is unaltered during moderate central hypoxemia (PaO₂ 50 mm Hg) [9]. On the other hand, hypoxia influences the rate of synthesis or release of neurotransmitters and modulators [12, 13], leading to accumulation of adenosine, dopamine and γ -aminobutyric acid in the brain [2, 14, 15]. Administration of long-acting analogues of adenosine, either systemically or directly into the third cerebral ventricle, decreases phrenic nerve activity in peripherally chemodenervated air breathing cats while theophylline, a specific antagonist of adenosine, prevents and reverses the decrease of phrenic activity [16] as reported by Javaheri et al. [17]. These studies [16, 17] clearly show the important role of adenosine in the central depressant effect of hypoxia in respiration in chemodenervated animals.

Adenosine is thought to be responsible for the depression of ventilation in chemodenervated animals during hypoxia. However, the cause of the secondary decrease of ventilation, which occurs in subjects with intact peripheral chemoreceptors during sustained moderate isocapnic hypoxia, has not been clarified. The neurotransmitters and neuromodulators that accumulate in the brain during hypoxia are thought to be responsible for the secondary decrease, but the particular neurotransmitter is not known. Adenosine, which is present in the brain in low concentration during normoxia and that increases during hypoxia, may also be responsible for this secondary decrease of ventilation in subjects with intact peripheral chemoreceptors during sustained moderate isocapnic hypoxia.

Therefore, in this study, the respiratory effects of theophylline, a central and peripheral adenosine receptor antagonist, on isocapnic moderate hypoxia in anesthetized cats were compared to those in a control group.

Materials and Methods

The experiments were conducted in cats anesthetized with Pentothal-Na (30 mg kg⁻¹ i.p.). A tracheotomy was done and a cannula connected to an inspiratory-expiratory valve was inserted into the trachea. Catheters were inserted into both femoral arteries for obtaining blood samples and for recording the systemic arterial pressure. The femoral vein was catheterized for administration of doses of anesthetic when necessary.

Fifteen cats were used as a control group. Eleven cats were injected with a specific adenosine antagonist (theophylline 13.6 mg kg⁻¹ i.v.). In order to eliminate its peripheral effects, in 5 cats theophylline (0.51 mg kg⁻¹) was injected into the cerebrospinal fluid (i.c.) by atlanto-occipital puncture over a 1-min period. All experimental animals were allowed to breathe either atmospheric air or 14% O₂-N₂ from the spirometer for 20 min.

Tidal volume (V_T), respiratory frequency (f) and arterial blood pressure were recorded while animals were breathing air and a hypoxic gas mixture. PaO₂ and PaCO₂ were determined. In order to maintain isocapnia, fractions of 1% CO₂ were added to the spirometer. PaCO₂ was maintained at the baseline value recorded before the prehypoxic phase. Systemic arterial pressure was recorded using a Statham pressure transducer. PaO₂ and PaCO₂ were determined using a AVL Gas check analyzer. Respiratory minute volume (VE) was calculated from V_T and f values. V_T and f were recorded by means of a pneumotachograph and a PT5 volumetric pressure transducer on a Grass 7 polygraph.

Results were statistically analyzed with a repeated-measures ANOVA. For adjustment of multiple comparisons, least significant difference was used ($p < 0.05$ was taken as statistically significant).

Results

When control animals were allowed to breathe the hypoxic gas mixture, V_T and VE increased significantly at 5 min (table 1). After this time, the magnitude of the increase in V_T diminished gradually. At 10–15 min after the onset of isocapnic hypoxia, V_T and VE values were not significantly different from the prehypoxic air phase. At 20 min after the onset of isocapnic hypoxia, VE was found to be significantly ($p < 0.05$) lower than that observed during the prehypoxic air phase (table 1).

PaCO₂ of the control animals was 34.7 ± 0.8 during the prehypoxic air phase and was kept within ± 2 mm Hg of control while animals were breathing the hypoxic gas mixture (table 1). The mean values of the systemic blood arterial pressure during the air breathing phase and 5 min after hypoxia were 162.2 ± 2.5 and 169.3 ± 1.9 mm

Table 1. Ventilatory and blood gas determinations for control and theophylline-injected (i.v. and i.c.) groups during air and isocapnic hypoxic phases (means \pm SE)

		f, min ⁻¹	V _T , ml	VE, ml	PaO ₂ , mm Hg	PaCO ₂ , mm Hg
Control (n = 15)	air	24.6 \pm 1.5	25.9 \pm 3.8	605.6 \pm 70.2	97.3 \pm 2.1	34.7 \pm 0.8
	5-min hypoxia	25.5 \pm 1.6	36.6 \pm 4.3***	895.5 \pm 85.2***	66.8 \pm 1.3***	33.4 \pm 0.6
	10-min hypoxia	23.9 \pm 1.1	22.4 \pm 2.3	509.2 \pm 58.7	68.6 \pm 1.0***	32.5 \pm 1.0
	15-min hypoxia	24.7 \pm 1.1	19.1 \pm 3.2	458.2 \pm 7.2	65.5 \pm 2.0***	32.4 \pm 0.8
	20-min hypoxia	23.9 \pm 1.1	22.2 \pm 3.3	465.2 \pm 52.6*	63.8 \pm 4.5***	32.0 \pm 1
i.v. theophylline (n = 11)	air	25.9 \pm 1.5	39.4 \pm 3.0	995.9 \pm 74.8	99.3 \pm 2.5	32.3 \pm 0.5
	5-min hypoxia	27.7 \pm 1.5**	49.8 \pm 3.1***	1,374.1 \pm 107.9***	65.9 \pm 1.1***	32.3 \pm 0.5
	10-min hypoxia	28.1 \pm 1.6**	48.5 \pm 3.7**	1,358.4 \pm 125.9**	66 \pm 1.1***	33.5 \pm 1.0
	15-min hypoxia	28.7 \pm 1.8*	48.4 \pm 4.4*	1,359.4 \pm 128**	65.1 \pm 2.0***	33.2 \pm 1.2
	20-min hypoxia	29 \pm 1.7**	49.8 \pm 3.5**	1,418 \pm 110.3***	66 \pm 1.2***	33.3 \pm 1.7
i.c. theophylline (n = 5)	air	21.6 \pm 1.2	32.7 \pm 1.0	705.2 \pm 44.9	99.4 \pm 3.1	39.2 \pm 0.9
	5-min hypoxia	22.2 \pm 1.3	40.6 \pm 1.2**	897.3 \pm 45.3**	68.7 \pm 1***	36.7 \pm 0.6
	10-min hypoxia	22 \pm 1.2	41.4 \pm 0.6**	909.2 \pm 46.9**	65.6 \pm 1***	36.5 \pm 0.9
	15-min hypoxia	22.4 \pm 0.9	40.6 \pm 1.1*	912.2 \pm 51.7*	65.6 \pm 0.8***	37.6 \pm 0.7
	20-min hypoxia	22.2 \pm 1.1	41.1 \pm 1.1**	911.8 \pm 41.6*	66.8 \pm 1***	38.1 \pm 1.0

f = Respiratory frequency; V_T = tidal volume; VE = ventilatory minute volume. Asterisks indicate statistical significance when compared with the prehypoxic air phase: * p < 0.05, ** p < 0.01, *** p < 0.001.

Table 2. Mean arterial blood pressure determination for control, i.v. theophylline and i.c. theophylline groups during air and isocapnic hypoxic phases (means \pm SE)

	Mean arterial blood pressure, mm Hg				
	air breathing	5-min hypoxia	10-min hypoxia	15-min hypoxia	20-min hypoxia
Control (n = 15)	162.2 \pm 2.5	169.3 \pm 1.9***	161.8 \pm 8.8	162.1 \pm 8.3	162 \pm 7.5
i.v. theophylline (n = 11)	127.4 \pm 5.7	128.6 \pm 5.4	126.2 \pm 6.2	120 \pm 7.1	120 \pm 7.1
i.c. theophylline (n = 5)	130 \pm 9.3	131 \pm 10.2	129 \pm 10.9	126.5 \pm 10.9	124 \pm 10.4*

Asterisks indicate statistical significance when compared with the prehypoxic air phase: * p < 0.05, *** p < 0.001.

Hg. Both paired t test and repeated-measures ANOVA showed that the difference was statistically significant (p < 0.001). At 10, 15 and 20 min of hypoxia, the systemic arterial pressure values were not different from those observed during the prehypoxic air phase (table 2).

In cats with i.v. injections of theophylline, f, V_T and VE increased significantly (p < 0.01, p < 0.001, p < 0.001) at 5 min of hypoxia. No decrease in the magnitude of the ventilatory response was observed during the 20-min period of hypoxic gas mixture breathing. At 20 min after the onset of hypoxia, f, V_T and VE values were significant-

ly (p < 0.01, p < 0.01, p < 0.001) greater than observed during the prehypoxic air phase (table 1). In cats injected with theophylline (i.c.), V_T and VE increased significantly (p < 0.01, p < 0.01) at 5 min of hypoxia. After this time, no decline in the magnitude of the response occurred during isocapnic hypoxia (table 1). In both theophylline-injected groups (i.v. and i.c.) no significant increase was observed in systemic arterial pressure during isocapnic hypoxia (table 2).

Discussion

Our results show that sustained (20-min) isocapnic moderate hypoxia causes an initial increase in ventilation followed by a decrease in respiratory minute volume. The initial increase in ventilation is brought about by the stimulation of peripheral chemoreceptors. Previous studies have demonstrated that the peripheral chemoreceptors are stimulated in acute hypoxia and in sustained hypoxia [4, 18]. The subsequent decline in ventilation (\dot{V}_E) that we observed was due to a decrease in V_T . No change was observed in respiratory frequency. The peripheral chemoreceptors have been shown to be active during ventilatory decline [4]. Vizek et al. [4] observed no decrement in carotid sinus nerve activity during ventilatory depression in sustained hypoxia. However, they observed a decrease in phrenic nerve activity [4]. These findings indicate that central mechanisms are responsible for the subsequent decrease of ventilation in sustained hypoxia. The central depressant effect of hypoxia on ventilation and particularly on V_T has long been established in chemodenervated and chemodenervated vagotomized animals [6, 8, 11]. The decrease in ventilation of chemodenervated animals during hypoxia was attributed to an increase in cerebral blood flow and the consequent decrease in cerebrospinal fluid PCO_2 [11] and to the release of inhibitory neurotransmitters [12, 13, 16]. However, studies by Javaheri et al. [17] on chemodenervated-vagotomized animals have shown that hypoxic ventilatory depression occurs both in the face of a rise or a fall in ventral medullary extracellular fluid PCO_2 and H^+ and is prevented by aminophylline.

In our present study V_T and V_E values of the theophylline-treated groups were higher than those of the control group during the air phase. This observation supports the findings of Eldridge et al. [19]. The results of these investigators showed that theophylline increases the respiratory drive by means of central neural mechanisms. Adenosine, which is present in the brain in low concentration, is known to inhibit the spontaneous neural activity in various parts of the brain. Theophylline, on the other hand, blocks the effects of adenosine by binding to the adenosine receptors. It is for this reason that V_T and V_E values of the theophylline-treated groups can be expected to be higher than those of the untreated group. When theophylline-treated groups were exposed to sustained moderate isocapnic hypoxia, respiration (V_T , \dot{V}_E) increased significantly in the first 5 min and remained at an increased level during the entire 20-min phase of hypoxic gas breathing. Secondary depression of ventilation was not observed in theophylline-treated groups. In other words, the subse-

quent decrease of ventilation that we observed in cats with intact peripheral chemoreceptors and vagi was absent in the i.v. and i.c. theophylline (a specific adenosine antagonist) groups. This finding indicates that adenosine, which accumulates in the brain during hypoxia, inhibits the central mechanisms and prevents their response to peripheral chemoreceptor impulses.

In the present study, systemic arterial pressure of the control cats was found to increase significantly at 5 min of hypoxia. This increase is due to peripheral chemoreceptor impulses impinging on the vasomotor area. The subsequent decrease in systemic arterial pressure observed after this time may be attributed to accumulation of neurochemical substances that inhibit pressor activity. It may also be due to peripheral effects of hypoxia on the cardiovascular system.

Arterial blood pressure was lower in theophylline groups (i.v. and i.c.). In the i.v. theophylline group this may be due to the systemic vasodilator effect of theophylline [20]. However, blood pressure was also low when theophylline was given i.c., therefore, we suggest a central effect of theophylline on vagal and vasomotor centers in the brain stem. Furthermore, the possibility of adenosine as a mediator cannot be excluded. There is experimental evidence that selective activation of A(1) adenosine receptors in the subpostremal nucleus tractus solitarii increases mean arterial pressure and causes an increase in preganglionic adrenal sympathetic activity [21]. The low blood pressure in our i.c. theophylline (adenosine antagonist) group may be due to inactivation of A(1) adenosine receptors in the nucleus tractus solitarii.

Conclusion

The results of this study show that accumulation of adenosine in the brain during sustained isocapnic moderate hypoxia seems to reduce the response of the central respiratory control mechanisms to chemoreceptor impulses and to cause the subsequent decrease of ventilation. Adenosine antagonist (theophylline) restores the response of central mechanisms to chemoreceptor impulses and prevents the secondary decline of ventilation.

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